

# A SELECTIVE METHOD FOR THE DEMONSTRATION OF BIFID BACTERIA (*LACTOBACILLUS BIFIDUS*) IN MATERIALS TESTED FOR FAECAL CONTAMINATION

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In the search for new and, if possible, more specific bacterial indicators of faecal contamination in foods and water, attention has recently been paid also to the Gram positive, anaerobic intestinal bacteria which mostly are referred to as *Lactobacillus bifidus* in the literature (5). Organisms of this type constitute the major part of the intestinal vegetation of milk-fed infants, but they are regularly encountered in high numbers also in the faeces of adults (e.g. ref. 4). DEHNERT (1) has described 4 different subtypes in this group. These subtypes can be distinguished on the basis of fermentation reactions and basic nutritional demands (1, 3; cf. Tables 1 and 2), and it seems that some of them are specific for certain habitats (2, 3, 7). The subtype occurring regularly in the faeces of human adults is referred to as type B<sub>1</sub> in tables 1 and 2, and it seems most convenient, therefore, to select this subtype as an indicator in the demonstration of faecal contamination in foods and/or water samples on the basis of the occurrence of bifid bacteria. A selective method for the enumeration of type B<sub>1</sub>, including the use of a simple synthetic medium, would therefore be very useful in investigations of this kind.

Table 1. Characteristic fermentation reactions of the different subtypes of bifid bacteria (+ indicates fermentation, 0 no fermentation).

Subtypes	Fermentation of					
	Glucose	Lactose	Xylose	Arabinose	Melezitose	Sorbitol
A	+	+	+	0	0	0
B <sub>1</sub>	+	+	+	+	+	0
B <sub>2</sub>	+	+	0	0	+	+
C	+	+	0	0	0	0

Table 2. Basic nutritional demands of the different subtypes of bifid bacteria (+ indicates essential nature of the given substance, s stimulatory effect, and 0 no dependence). The following abbreviations are employed: B = biotine, Pa = pantothenic acid, P = pantethine, R = riboflavine, Pep = peptide factor, As = amino sugar factor, PPB = Mixture of purine and pyrimidine bases, Oa = oleic acid, C = cystine.

Subtypes	B	Pa	P	R	Pep	As	PPB	Oa	C
A	+	+	0	0	0	0	0	s	+
B <sub>1</sub>	s		+	+	s	0	0	s	+
B <sub>2</sub>	s		+	+	s	0	0	s	+
C	s		+	+	+/s	+	+	s	+

### *Description of the method*

A simple medium for the enumeration of type A which predominates in breast-fed infants has been developed by PETUELY (6,8). As shown in Table 2 the nutritional demands of type B<sub>1</sub> are somewhat more complex than those of type A. The difference is that type B<sub>1</sub> requires riboflavine, and most strains also preformed pantethine. The medium of PETUELY must thus be modified so as to contain also riboflavine and pantethine in order to satisfy the requirements of type B<sub>1</sub>. Since type A, on the other hand, fails to utilize arabinose, the medium becomes selective for type B<sub>1</sub> if arabinose replaces lactose as the source of energy. The medium for a selective enumeration of has the following composition.

L(+)-arabinose	1 g	FeSO <sub>4</sub> ·7H <sub>2</sub> O	1 mg
Cysteine-HCl	0.04 g	MnSO <sub>4</sub> ·4H <sub>2</sub> O	0.7 mg
K <sub>2</sub> HPO <sub>4</sub>	0.5 g	NaCl	1 mg
(NH <sub>4</sub> ) <sub>2</sub> SO <sub>4</sub>	0.4 g	Sodium ascorbate (in terms of ascorbic acid)	1 g
Tween 80	0.1 g	Washed agar	2 g
Pantethine	50 μg	Distilled water	ad 100 ml
Riboflavine	25 μg	pH is adjusted to 6.8—7.0	
Biotine	0.5 μg		
MgSO <sub>4</sub> ·7H <sub>2</sub> O	20 mg		

This medium is prepared in two lots, one of which contains all the components except arabinose and ascorbic acid at double strength. This lot can be sterilized by ordinary autoclaving. The other lot is prepared by dissolving 1 g ascorbic acid in distilled water; 1-N NaOH is added to bring the pH into the range of 6.8—7.0. 1 g arabinose is then dissolved in this Na-ascorbinate solution, and distilled water is added to bring the volume to 50 ml. This solution is sterilized by filtration. Equal volumes of the ascorbate-arabinose solution (heated to 55—60°) and the melted agar lot (cooled to 55—60°) are then mixed, the mixture is poured on plates and allowed to solidify. Overheating of the ascorbate-arabinose solution must be avoided since it gives rise to oxidation of the ascorbic acid.

The plates of the final medium are inoculated for the enumeration of type B<sub>1</sub> with suitable dilutions of the material to be tested by means of the »surface drop»

method. The testing of the water samples can be performed with the membrane filter method. In order to facilitate the counting of the colonies from the membranes we have used an 0.01 per cent concentration of 2, 3, 5-triphenyltetrazolium chloride (referred to below as TTC) in the medium. TTC was added to the ascorbate-arabinose solution immediately before filtration. In the presence of TTC, overheating of the solution results in the spontaneous precipitation of TTC as formazan, the critical point seems to be as low as about 40° C.

Incubation is performed at 37° C in carbon dioxide for 48 hours. For this purpose ordinary laboratory desiccators can be employed. They are emptied to about 10—15 mm Hg with a water pump, and are then filled with commercial CO<sub>2</sub>. Under these conditions type B<sub>1</sub> grows on the synthetic agar in the form of smooth, circular and convex colonies. They are almost colorless, have entire margins, and are 0.2—2 mm in diameter. In the presence of TTC flakes of red precipitate appear in the colonies, which are thus easily distinguished from the background (Figure 1).

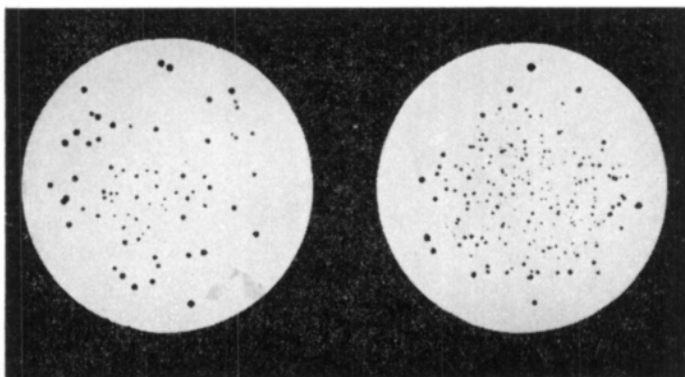


Figure 1. Colonies of type B<sub>1</sub> on membranes incubated on selective synthetic agar containing TTC.

*Experiences gained from of the method in use.*

We have employed the method described above for testing faecal samples of human (adult) origin, samples of water artificially polluted with faeces, and ordinary water samples. A considerable number of the colonies which grew on the synthetic medium in the experiments were examined microscopically, and it was found that only less than 1 per cent of the colonies were representative of organisms other than bifid bacteria. In the concentration of TTC employed by us a slight reduction (15—40 per cent) in the numbers of type B<sub>1</sub> colonies was observed. Since the type B<sub>1</sub> figures nevertheless remained 10—100 times higher than those of coliform organisms (Congo agar) or enterococci (M Enterococcus agar; 9) determined from the same samples, it appears that the addition of TTC does not effect a considerable loss in the relative sensitivity of the method. The significance of type B<sub>1</sub> in foods and water is still to be assessed accurately, but for the present it can be concluded that the method described here is highly selective for type B<sub>1</sub> which is a typical intestinal organism, though hitherto not proved as occurring regularly in other environments.

## Summary

A method which permits a selective enumeration of a subtype of the organisms hitherto referred to as *Lactobacillus bifidus*, known as an ordinary inhabitant of the intestines of human adults, is described. It is suggested that this method may be useful for the demonstration of organisms of this kind in foods and water as indicators of faecal pollution.

### REFERENCES.

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### SELOSTUS:

VALIKOIVA MENETELMÄ »BIFIDIBAKTEERIEN» (*LACTOBACILLUS BIFIDUS*)  
OSOITTAMISEKSI FEKAALISEN SAASTUMISEN SUHTEEN TUTKITUISTA  
NÄYTTEISTÄ

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Esitetään menetelmä, jonka avulla erään *Lactobacillus bifiduksen* alatyypin valikoiva viljeleminen on mahdollista. Koska ko. bakteeria aina tavataan varsin runsaasti aikuisten ulosteissa on todennäköistä, että sen esiintyminen esimerkiksi vesi- ja elintarvikenäytteissä on fekaalisen kontaminaation herkkä indikaattori. Nyt kuvattu menetelmä näyttäisi siten soveltuvan tällaisen kontaminaation osoittamiseen.