

Symptom expression and accumulation of potato virus Y (PVY^O) and potato leaf roll virus in thirteen potato cultivars

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Necrotic local lesions developed in cvs. Matilda, Ostara, Record, Saturna, Stina, Hankija's (Hjan) Tanu and Hjan Timo and local ring spots in Olympia and Sieglinde (Siikli) following sap inoculation with the ordinary strain of potato virus Y (PVY^O). Secondarily infected cvs. Ostara, Pito, Siikli and Hjan Timo developed leaf drop. No infected progeny was produced by Matilda, Saturna and Hjan Tanu. In contrast, Bintje, Puikula and Sabina developed neither local lesions nor systemic necrosis, but showed mosaic symptoms following primary and secondary infection by PVY^O. The ELISA absorbance values for potato leaf roll virus (PLRV) in Ostara, Pito and Saturna were less than 10% of those in the PLRV-infected Siikli. The ELISA values for PLRV in Olympia, Stina, Hjan Tanu and Hjan Timo were not significantly different from those of Siikli. The severity of the symptoms did not correlate with the concentration of PLRV in the potatoes.

Key words: hypersensitivity, virus accumulation, virus resistance, potato

Introduction

Potato virus Y (PVY) and potato leaf roll virus (PLRV) are the economically most important viruses of potato (*Solanum tuberosum* L.) in Europe, causing yield losses of up to 80% (DE BOKX and van der WANT 1987). In Scandinavia, PVY is more important than PLRV (UMAERUS et al. 1979, KURPPA 1983). In Finland, the tobacco vein necrosis strain of PVY (PVY^N) seems to be dominating, whereas the ordinary strain of PVY (PVY^O) is the one most commonly encountered in other parts of Europe (KURPPA 1983, DE BOKX and van der WANT 1987).

Resistance in potato to viruses is important in the control of viral diseases. The types of resistance to PVY are hypersensitivity and extreme resistance

(ROSS 1986). Localized hypersensitivity is expressed as necrotic local lesions in the sap-inoculated leaves. Systemic hypersensitivity results in necrosis in the top or the top leaves of the PVY-infected plant. ROSS (1986) defined extreme resistance as intensified local hypersensitivity. We define extreme resistance as the type where the virus concentration remains extremely low in an infected plant. The latter definition not only includes the type of resistance in which the multiplication of PVY is reduced (BARKER and HARRISON 1984), but also the possibility that the low virus concentration in plant tissue is due to the inhibited cell-to-cell spread of the virus (VALKONEN et al. 1991).

The three components of resistance to PLRV in potato are restriction of virus multiplication, resistance to infection and inhibition of virus movement

from foliage to tubers (BARKER and HARRISON 1985, 1986, BARKER 1987). The first component was considered to be the most important by BARKER and SOLOMON (1990), who detected restriction of PLRV accumulation in some cultivated potato genotypes. Furthermore, extreme resistance to PLRV has been found in a few wild potato species (JONES 1979, BROWN et al. 1984, BROWN 1991, VALKONEN et al. 1992).

The local potato production in Finland is mainly based on Finnish, Swedish, German and Dutch cultivars. However, the information about the type and level of resistance to PVY and PLRV in the potato cultivars commonly grown in Finland is limited and data has been obtained mostly from the field and not from experiments performed under controlled conditions (KURPPA 1983, STEGEMANN and SCHNICK 1985, KURPPA and HASSI 1989). Determination of the type and level of virus resistance in the above potato cultivars was considered important for the strategic planning of the potato breeding programme for virus resistance initiated recently as a joint project of the Department of Plant Production, University of Helsinki, and the Institute of Plant Breeding, Agricultural Research Centre of Finland. The present study was undertaken to test the symptom expression and accumulation of PVY^O and PLRV in 13 potato cultivars grown in Finland as PVY^O and PLRV are the most important viruses of potato worldwide, PVY^O usually causes more severe symptoms in potato than does PVY^N (DE BOKX and van der WANT 1987), and PLRV may become more important also in Scandinavia as a consequence of the forecasted warming up of the climate (CARTER 1992).

Material and methods

Viruses

One isolate each of PLRV and PVY^O was obtained from fieldgrown potatoes in Sudan and England, respectively (EL-AMIN et al. 1990, GIBSON et al.

1990). These virus isolates were used, because the above isolate of PVY^O is well-characterized (GIBSON et al. 1990, VALKONEN et al. 1991, 1992) and resembles the Finnish PVY^O isolate YSF11 of KURPPA (1983) in terms of biological properties, and because PLRV occurs only sporadically in Finnish potato fields and no local isolates of the virus were available. PVY^O was maintained in *Nicotiana tabacum* cv. Samsun and PLRV in potato cv. Sieglinde.

Antisera

The antibodies and alkaline phosphatase conjugated antibodies to PVY and PLRV were obtained from Böhringer.

Plants and growing conditions

Virus-tested seed potatoes of cvs. Bintje, Ostara, Record and Saturna (Dutch), Olympia and Sieglinde (Siikli) (German), Matilda, Sabina and Stina (Swedish), and Pito, Puikula, Hankkija's (Hjan) Tanu and Hjan Timo (Finnish) (STEGEMANN and SCHNICK 1985) were obtained from the Finnish Seed Testing Institute, Helsinki. The tubers were planted into pots 21 cm in diameter and filled with a mixture of steam-sterilized peat and washed sand (10:1 v/v) in the beginning of July 1991, and the pots were then sunk by halfway into a sand bed in an aphid-proof nethouse. The plants were watered daily and fertilized weekly with NPK fertiliser. Late blight (*Phytophthora infestans* (Mont.) de Bary) was controlled by applications of methalaxyl, mancozeb and maneb. The average photoperiod was 16h and the mean temperatures at day and night were 26°C and 16°C, respectively. The tubers were harvested at the end of September and dried at room temperature for two days. After storage of four months at 4°C, five randomly taken progeny tubers per plant were planted into pots and grown in glasshouse as described above. The photoperiod was extended to 18 h by illumination with fluorescent lamps. The mean temperatures during day and night were as above.

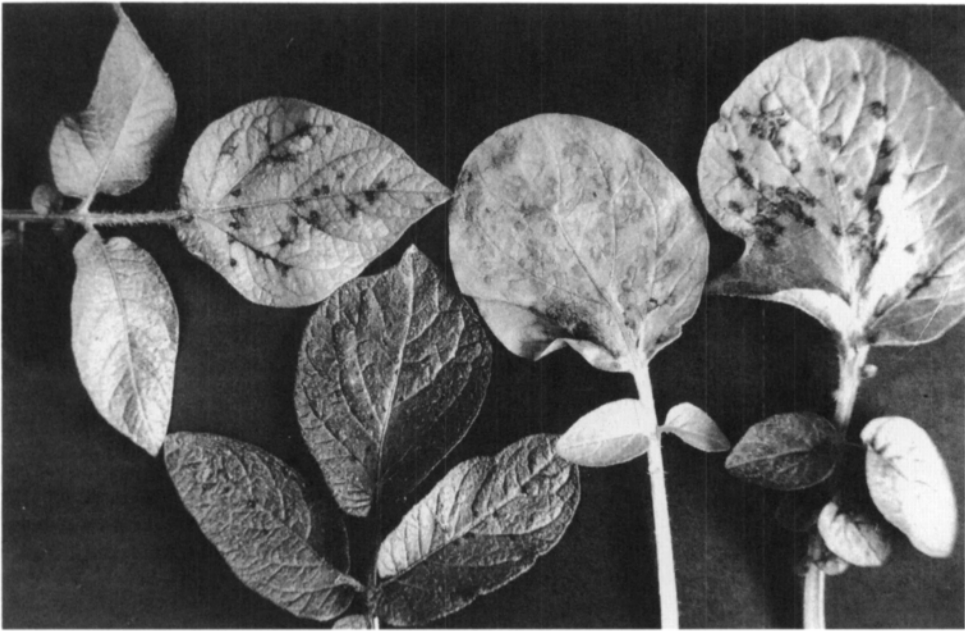


Fig. 1. Local lesions following sap inoculation with PVY^O. From the left: cvs. Matilda, Record, Siikli and Stina.

Virus inoculation

PVY^O was sap-inoculated by grinding leaves of infected tobacco at 1g/5ml of distilled water with a pestle and mortar, and rubbing the extract onto carborundum-dusted leaves of the potato plants. Two plants per potato cultivar were inoculated with PVY^O. The two oldest leaves of four shoots per plant were inoculated at the emergence of the fifth leaf of the shoot (stage of development no. 305) (JEFFERIES and LAWSON 1991), whereas all other shoots were cut off.

PLRV was graft-inoculated, which is the most sensitive method of testing the resistance to PLRV (SWIEZYNSKI et al. 1989). Two plants per cultivar were inoculated at the emergence of the seventh leaf of the shoot (stage of development no. 307) (JEFFERIES and LAWSON 1991). Four shoots per plant were left growing; all other shoots were cut off. Two of the four shoots were then graft-inoculated with PLRV by removing a leaf from the upper

part of the stem with its axillary bud, cutting the stem lengthways, sidegrafting an infected scion of cv. Siikli in place of the removed bud and binding it with Parafilm. The growth of the infected scion was taken as an indication of a successful graft union and the scions were removed three weeks after grafting.

Virus detection

The local and systemic symptoms were visually observed 14 and 28 days after sap inoculation of PVY^O. The secondary symptoms were visually observed and the titres of PVY^O and PLRV were determined by a double antibody sandwich enzyme-linked immunosorbent assay (DAS-ELISA) (CLARK 1981) in the progeny plants at the appearance of flower buds (stage of development no. 410) (JEFFERIES and LAWSON 1991). For DAS-ELISA, the uppermost fully expanded leaves of the plants

Table 1. Symptoms due to primary infection (sap-inoculated PVY^O), and symptoms and ELISA absorbance values (A₄₀₅) ± standard deviation due to secondary infection by PVY^O, as determined at the time of flower initiation.

| | Primary infection symptoms | | Secondary infection | | |
|--------------------------|----------------------------|----------|------------------------|----------|-------------------------|
| | local | systemic | no. of tubers infected | symptoms | A ₄₀₅ values |
| Bintje | O | M | 5/5 | M, S, R | 1.61 ± 0.13 |
| Matilda | NRS, VN | VN, IVN | 0/5 | O | 0.02 ± 0.00 |
| Olympia | RS | VN, CS | 4/5 | M | 1.36 ± 0.22 |
| Ostara | NS | O | 2/5 | M, LD | 1.61 ± 0.17 |
| Pito | O | VN, CS | 5/5 | M, LD | 1.72 ± 0.11 |
| Puikula | O | M | 4/5 | M, R | 1.72 ± 0.13 |
| Record | NS | O | 3/5 | M | 1.45 ± 0.18 |
| Sabina | O | M | 5/5 | M | 1.55 ± 0.30 |
| Saturna | NRS, VN | VN | 0/5 | O | 0.03 ± 0.01 |
| Siikli | RS | NRS, VN | 2/5 | M, LD | 1.07 ± 0.50 |
| Stina | NRS, VN | VN, IVN | 1/5 | M, LD | 0.28 |
| Hjan Tanu | NRS | O | 0/5 | O | 0.03 ± 0.01 |
| Hjan Timo | NRS | O | 4/5 | M, LD | 1.32 ± 0.29 |
| non-inoculated cultivars | | | | | 0.02 ± 0.01 |

O = no symptoms
 CS = chlorotic spots
 IVN = interveinal necrosis
 LD = leaf-drop
 M = mosaic
 NRS = necrotic ringspots
 NS = necrotic spots
 R = rugosity
 S = stunting
 VN = vein necrosis
 RS = ringspots

were sampled in duplicate, extracted at 1g/3ml of buffer and pipetted onto ELISA plates (Greiner Labortechnik) in aliquots of 200 µl. Absorbances were recorded at 405 nm (A₄₀₅) after developing the colour reaction for 1 h using p-nitrophenyl as substrate.

Statistical analysis

Analysis of variance was used for the statistical analysis of the A₄₀₅ values and calculations of the least significant differences (LSD) when appropriate (STEEL and TORRIE 1981).

Results

PVY^O

Necrotic local lesions were observed in cvs. Matilda, Ostara, Record, Saturna, Stina, Hjan Tanu and Hjan Timo following sap inoculation with PVY^O (Table 1), whereas Olympia and Siikli developed ring spots that remained green while the leaves turned yellow (Fig. 1). Bintje, Pito, Puikula and Sabina showed no symptoms in the sap-inoculated leaves.

Systemic vein necrosis following primary infection was observed in Matilda, Olympia, Pito, Sa-

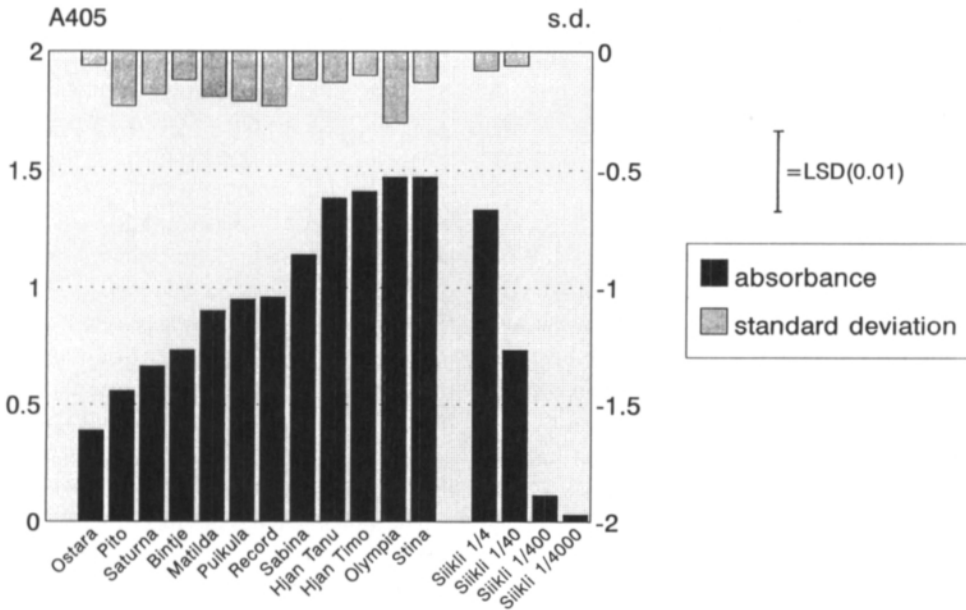


Fig. 2. Absorbance values (A_{405}) (ELISA) and their least significant differences at a risk level of 1% ($LSD_{0.01}=0.34$) for detection of PLRV in the foliage of the progeny of potato cultivars graft-inoculated with PLRV. The dilution of the sap from cv. Siikli was 4 - 4000-fold and that of the other cultivars 4-fold. A_{405} values for uninfected cultivars were similar to those for the 4000-fold dilution of the sap of PLRV-infected cv. Siikli.

turna, Siikli and Stina (Table 1). Systemic mosaic symptoms without necrosis were observed in Bintje, Puikula and Sabina, while Ostara, Record, Hjan Tanu and Hjan Timo were free of any.

Systemic mosaic symptoms were observed in the progeny plants of ten of the 13 cultivars following secondary infection by PVY^O (Table 1). Furthermore, Ostara, Pito, Siikli and Hjan Timo developed leaf drop, Bintje was heavily stunted and the growth of Puikula was poor. In contrast, the progeny of Matilda, Saturna, Stina and Hjan Tanu were free of symptoms and PVY^O according to ELISA.

PLRV

All the tested progeny tubers of each cultivar graft-inoculated with PLRV were infected with PLRV according to ELISA. However, the cultivars fell into three groups according to the PLRV titres ($P=0.01$) (Fig. 2). Olympia, Siikli, Stina, Hjan Tanu and Hjan Timo yielded high PLRV titres ($A_{405} >$

1.33), Matilda, Puikula and Record yielded moderate PLRV titres ($0.90 < A_{405} < 0.96$), and Ostara, Pito and Saturna yielded low PLRV titres ($A_{405} < 0.66$). The PLRV titres of Bintje ranged between low and moderate and those of Sabina between moderate and high ($LSD_{0.01}=0.34$ for the A_{405} values).

PLRV caused yellowing of the leaves and leaf roll in Matilda, Ostara, Siikli and Hjan Timo, yellowing of the leaves without leaf roll in Olympia, Puikula, Saturna and Hjan Tanu, and yellowing of the leaf margins in Pito. Symptomless infection by PLRV was detected in Bintje, Record, Sabina and Stina.

Discussion

Hypersensitivity reactions were observed in 10 of the 13 cultivars following infection with PVY^O. Expression of hypersensitivity is useful, as such a response induced by a virus or virus strain in a plant

can reduce the systemic spread of the virus (FRITIG et al. 1987). Indeed, no infected progeny was detected in Matilda, Saturna and Hjan Tanu and few infected progeny tubers were produced by Stina. These cultivars reacted by local necrosis to primary infection of PVY^O. Similar results were obtained in Matilda by KURPPA and HASSI (1989).

There are few studies on the genetic control of hypersensitivity to PVY in potato. COCKERHAM (1970) identified four genes which control the hypersensitivity to all strains of PVY in a number of *Solanum* spp., of which the genes in *S. chacoense* Bitt. and *S. demissum* Lindl. acted also against potato virus A (PVA). JONES (1990) suggested that the strain group specific hypersensitivity to PVY^O in certain potato genotypes was controlled by a single dominant gene, Ny_{thr}, which possibly originated in Katahdin and a Scottish potato clone 11-79 (DAVIDSON 1980), and by another dominant gene of unknown origin. Our results do not allow further comparisons of the genes for hypersensitivity to PVY between the cultivars of our study and those reported by COCKERHAM (1970) and JONES (1990), as those comparisons would require inoculations with other strains of PVY and PVA. However, the hypersensitivity to PVY^O in Hjan Timo, a cultivar derived from the cross Frühnudel x Katahdin (VARIS 1975), is presumably controlled by the gene Ny_{thr} from Katahdin.

The progenies of Bintje, Puikula and Sabina were almost 100% infected with PVY^O. The plants generated high PVY titres and showed severe mosaic symptoms without necrosis. The susceptibility of these cultivars to PVY hampers their cultivation in Southern Scandinavia. Therefore, Puikula and Sabina are mainly grown in the northernmost areas, where the spread of PVY is reduced due to the low populations of the PVY transmitting species of aphids (UMAERUS et al. 1979, SIGVALD 1984, KURPPA and RAJALA 1986).

The incidence of PLRV-infected progeny tubers was similarly high in all of the cultivars. However, there were variations in the PLRV concentrations between the progenies of cultivars. The concentration of PLRV in cultivars such as Ostara, Pito and Saturna was less than 10% of that in Siikli. Restricted accumulation of PLRV is useful in potato

as it potentially reduces the spread of PLRV by aphids (BARKER and HARRISON 1986).

The severity of the symptoms and the concentrations of PLRV did not strictly correlate in the potato genotypes tested. For example, apparent leaf roll and yellowing symptoms were observed in Ostara which exhibited the lowest PLRV concentration of all the cultivars. In contrast, no symptoms were observed in Stina which generated high concentrations of PLRV. The severity of symptoms observed in the PLRV-infected progeny of potato correlates rather with the physiological stage of the mother plants at the time of inoculation than with the PLRV concentrations in the progeny plants (BARKER and HARRISON 1986, BARKER and WOODFORD 1987).

The valuable types of virus resistance, i.e. hypersensitivity to PVY^O and restricted accumulation of PLRV, were incorporated in e.g. Ostara and Saturna. However, no cultivar exhibited extreme resistance to PVY and PLRV. Genes for extreme resistance are the obvious choice to be incorporated in new cultivars (JONES 1990), because extreme resistance acts against most, if not all strains of a virus and efficiently reduces the virus transmission by aphids (JONES 1979, 1990, GIBSON et al. 1990). Furthermore, virus strains capable of overcoming extreme resistance are not common in the field (JONES 1985, ROSS 1986). Extreme resistance to PVY, PLRV and PVA is incorporated in the wild potato species of the Etuberosa group, e.g. *S. brevifolius* Phil. (VALKONEN et al. 1992), and it has recently been transferred to some cultivated potato genotypes (PEHU et al. 1990, WILLIAMS et al. 1990, XU et al., unpublished results). The results suggest higher acceptability and productivity for most cultivars of the present study in Southern Scandinavia and other parts of Europe, where PLRV and PVY^O are the most disastrous viruses in potato, if genes for extreme resistance to PLRV and PVY could be incorporated into them.

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References

- BARKER, H. 1987. Multiple components of the resistance of potatoes to potato leaf roll virus. *Ann. Appl. Biol.* 111: 641-648.
- & HARRISON, B. D. 1984. Expression of genes for resistance to potato virus Y - potato plants and protoplasts. *Ann. App. Biol.* 105: 531-545.
- & HARRISON, B. D. 1985. Restricted multiplication of potato leaf roll virus in resistant potato genotypes. *Ann. App. Biol.* 107: 205-212.
- & HARRISON, B. D. 1986. Restricted distribution of potato leaf roll antigen in resistant potato genotypes and its effect on transmission of the virus by aphids. *Ann. Appl. Biol.* 109: 595-604.
- & SOLOMON, R. M. 1990. Evidence of simple genetic control in potato of ability to restrict potato leaf roll virus concentrations in leaves. *Theor. Appl. Genet.* 80: 188-192.
- & WOODFORD, J. A. T. 1987. Unusually mild symptoms of potato leafroll virus in the progeny of late-infected mother plants. *Potato Res.* 30: 345-348.
- BROWN, C. R. 1991. Extreme resistance to potato leaf roll virus from *Solanum chacoense*. 75th Ann. Meet. Potato Assoc. America. *Am. Potato J.* 68: 598.
- , SALAZAR, L., OCHOA, C., CHAVEZ, R., SCHILDE-RENSCHLER, L. & LIZARRAGA, C. 1984. Ploidy manipulation of a new source of resistance to PLRV from *Solanum acule*. 9th Trienn. Conf. Eur. Assoc. Potato Res. Interlaken, Switzerland. Abstr. 288-289.
- CARTER, T. 1992. The greenhouse effect and Finnish agriculture. In: *The First Hundred Years*. National Board of Agriculture. Helsinki, Finland. p. 31-57.
- CLARK, M. F. 1981. Immunoserbent assays in plant pathology. *Ann. Rev. Phytopathol.* 19: 83-106.
- COCKERHAM, G. 1970. Genetical studies on resistance to potato viruses X and Y. *Heredity* 25: 309-348.
- DAVIDSON, T. M. W., 1980. Breeding for resistance to virus disease of the potato (*Solanum tuberosum*) at the Scottish Plant Breeding Station. *Rep. Scott. Pl. Breed. Sta.* 1979. p. 100-108.
- DE BOKX, J. A. & WANT, J. P. H. van der 1987. *Viruses of Potatoes and Seed-Potato Production*. PUDOC, Wageningen. 259 p.
- EL-AMIN, S. M., BREMER, K., VARIS, E. & PEHU, E. 1990. Identification and eradication of potato viruses from an indigenous Sudanese potato stock (Zalinge). 11th Trienn. Conf. Eur. Assoc. Potato Res. Edinburgh, UK. Abstr. 72.
- FRITIG, G., KAUFFMANN, S., DUMAS, P., GEOFFROY, P., KOPP, M. & LEGRAND, M. 1987. Mechanism of hypersensitivity reaction of plants. In: *Plant Resistance to Viruses*. Ciba Foundation Symposium 133. Wiley, Chister. p. 92-108.
- GIBSON, R. W., PEHU, E., WOODS, R. D. & JONES, M. G. K. 1990. Resistance to potato virus Y and potato virus X in *Solanum brevidens*. *Ann. Appl. Biol.* 116: 151-156.
- JEFFERIES, R. A. & LAWSON, H. M. 1991. A key for the stages of development of potato (*Solanum tuberosum*). *Ann. Appl. Biol.* 119: 387-399.
- JONES, R. A. C. 1979. Resistance to potato leaf roll virus in *Solanum brevidens*. *Potato Res.* 22: 149-152.
- 1985. Further studies on resistance-breaking strains of potato virus X. *Pl. Pathol.* 34: 182-189.
- 1990. Strain group specific and virus specific hypersensitive reactions to infection with potyviruses in potato cultivars. *Ann. Appl. Biol.* 117: 93-105.
- KURPPA, A. 1983. Potato viruses in Finland and their identification. *J. Sci. Agric. Soc. Finl.* 55: 183-300.
- & HASSI, A. 1989. Reaction of four table potato cultivars to primary and secondary infection by potato viruses Y^O and Y^N. *Ann. Agric. Fenn.* 28: 297-307.
- KURPPA, S. & RAJALA, P. 1986. Occurrence of winged aphids on potato plants and pressure for potato virus Y transmission in Finland. *Ann. Agric. Fenn.* 25: 199-214.
- PEHU, E., GIBSON, R. W., JONES, M. G. K. & KARP, A. 1990. Studies on the genetic basis of resistance to potato leaf roll virus, potato virus Y and potato virus X in *Solanum brevidens* using somatic hybrids of *S. brevidens* and *S. tuberosum*. *Pl. Sci.* 69: 95-101.
- ROSS, H. 1986. Potato breeding - problems and perspectives. *Advances in Plant Breeding*. Supplement 13. Paul Parey, Berlin. p. 64-75.
- SIGVALD, R. 1984. The relative efficiency of some aphid species as vectors of potato virus Y^O (PVY^O). *Potato Res.* 27: 285-290.
- STEELE, R. G. D. & TORRIE, J. H. 1981. *Principles and Procedures of Statistics. A Biometrical Approach*. McGraw-Hill, New York. 633 p.
- STEGEMANN, H. & SCHNICK, D. 1985. Index 1985 of European potato varieties. *Mitteil. Biol. Bundesanstalt Land- und Forstwirts.* Heft 227. Berlin. 126 p.
- SWIEZYNSKI, K. M., DZIEWONSKA, M. A. & OOTROWSKA, K. 1989. Resistance to potato leaf roll virus (PLRV) in diploid potatoes. *Pl. Breed.* 103: 221-227.
- UMAERUS, M., HALLING, S. & RUUTH, P. 1979. Potatisförädling för norra Sverige. *Sveriges Utsädesförenings Tidskrift*. p. 47-66.
- VALKONEN, J. P. T., PEHU, E., JONES, M. G. K. & GIBSON, R. W. 1991. Resistance in *Solanum brevidens* to both potato virus Y and potato virus X may be associated with slow cell-to-cell spread. *J. Gen. Virol.* 72: 231-236.
- , BRIGNETI, G., SALAZAR, L. F., PEHU, E. & GIBSON, R. W. 1992. Interactions of the *Solanum* spp. of the Etuberosa group and nine potato-infecting viruses and a viroid. *Ann. Appl. Biol.* 120: 301-313.
- VARIS, E. 1975. Hankkija's Timo potato. Five-year Rep. Hankkija Pl. Breed. Inst. p. 163-166.
- WILLIAMS, C. E., HUNT, G. J. & HELGESON, J. P. 1990. Fertile somatic hybrids of *Solanum* species: RFLP analysis of a hybrid and its sexual progeny from crosses with potato. *Theor. Appl. Genet.* 80: 545-551.

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SELOSTUS

Perunan Y-viruksen ja perunan kierrelehtisyysviruksen oireet ja pitoisuus kolmessatoista perunalajikkeessa

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Perunan Y-virus (PVY) ja perunankierrelehtisyysvirus (PLRV) ovat haitallisimmat perunaa infektioivat virukset maailmanlaajuisesti, ja erityisesti PVY on merkittävä satotappioiden aiheuttaja Pohjoismaissa. Kolmentoista Suomessa yleisesti viljeltävän perunalajikkeen kestävyttä PVY:n yleistä rotua (PVY^O) ja PLRV:ta vastaan tutkittiin, sillä käytettävissä ei ollut riittävästi tietoa kyseisten lajikkeiden PVY- ja PLRV-kestävyydestä perunan virusresistenssijalostusohjelman suunnittelun perustaksi.

PVY:lla saastutettuihin lehtiin muodostui kuoliolaikkuja lajikkeissa Matilda, Ostara, Record, Saturna, Stina, Hankkijan (Hjan) Tanu ja Hjan Timo, kun taas Olympiassa ja Sieglindessa (Siikli) havaittiin rengaslaikkuja. Saastuttamattomissa ylälehdissä havaittiin (systeemistä) suonikuoliota lajikkeissa Matilda, Olympia, Pito, Saturna, Siikli ja Stina. Näissä lajikkeissa ilmeni N-geenin tuottamaa hypersensitiivisyyttä PVY^O:ta vastaan. Sen sijaan Bintjessä, Puikulassa ja Sabinassa ilmeni systeemistä viherkirjoa ilman kuoliota. PVY:n saastuttamista mukuloista kasvaneiden Ostaran, Pidon, Siiklin ja Hjan Timon taimien alalehdet kuolivat ja jäivät roikkumaan varresta. PVY:lla saastutettujen Matildan, Saturnan ja Hjan Tanun mukuloista yksikään ei tuottanut virussaastuneita taimia, kun taas muiden PVY:lla saastutettujen lajikkeiden mukuloista kasvaneissa taimissa ilmeni viherkirjoa, ja taimissa havaittiin korkeita PVY-pitoisuuksia

ELISA-testeissä.

Kaikki lajikkeet, jotka oli saastutettu PLRV:lla varttamalla PLRV-infektioitunut perunanverso tutkittavan lajikkeeseen, tuottivat PLRV-saastuneita mukuloita. Saastuneista mukuloista kasvaneiden taimien PLRV-pitoisuus vaihteli lajikkeittain siten, että se oli alhainen Ostarassa, Pidossa ja Saturnassa, keskinkertainen Matildassa, Puikulassa ja Recordissa, ja korkea Olympiassa, Siiklissä, Stinassa, Hjan Tanussa ja Hjan Timossa. Bintjen PLRV-pitoisuus sijoittui oli alhaisen ja keskinkertaisen välille ja Sabinan keskinkertaisen ja korkean välille. PLRV aiheutti lehtien kellastumista ja kiertymistä Matildassa, Ostarassa, Siiklissä ja Hjan Timossa, pelkästään lehtien kellastumista Olympiassa, Puikulassa, Saturnassa ja Hjan Tanussa ja lehtien reunojen kellastumista Pidossa. PLRV-saastuneessa Bintjessä, Recordissa, Sabinassa ja Stinassa ei havaittu oireita. Siten lajikkeiden PLRV-pitoisuuden ja oireiden voimakkuuden välillä ei ollut johdonmukaista riippuvuutta.

Ostarassa ja Saturnassa yhdistyivät yliherkkyys (hypersensitiivisyys) PVY^O:ta vastaan ja alhainen PLRV-pitoisuus, mikä voi merkittävästi ehkäistä PVY^O:n ja PLRV:n leviämistä näissä lajikkeissa. Sen sijaan missään lajikkeessa ei havaittu äärimmäistä R-geenin tuottamaa kestävyttä PVY:ta ja PLRV:ta vastaan.