

Fine structure and distribution of antennal sensilla of stink bug *Arma chinensis* (Heteroptera: Pentatomidae)

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Scanning electron microscopy was used to examine the morphology, ultra-structure, and distribution of antennal sensilla of the stink bug *Arma chinensis*. Two types of sensilla trichodea (ST 1–2), four types of sensilla basiconica (SB 1–4), one type of sensilla chaetica (SCH), one type of sensilla cavity (SCA) and one type of sensilla coeloconica (SCO) were distinguished on the antennae in both sexes. ST1 and ST2 were absent from the scape and pedicel. SB1 were absent from the scape. SB2 were distributed throughout the antennae. SB3 were located on the second pedicel and the two flagellomeres. SB4 were absent from the second flagellomere. SCH was observed on the second pedicel and the two flagellomeres. SCA and SCO occurred only on the second flagellomere. SB1 clusters occurred on the distal part of the second flagellomere. We compared the morphology and structure of these sensilla to other Heteroptera and discuss their possible functions.

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1. Introduction

Arma chinensis (Fallou) (Heteroptera: Pentatomidae) is an important predaceous insect species that is widely distributed throughout eastern Asia. It is an important and a broadly-applied biological control agent of forest pests in China. They are usually found on elm and poplar, and in cotton and soybean fields (Xu *et al.* 1981, Song *et al.* 2010, Zou *et al.* 2012). They prey upon a large

variety of species, and can effectively suppress agricultural and forest pests in the orders Lepidoptera, Coleoptera, Hymenoptera and Hemiptera (Xu *et al.* 1984, Zou *et al.* 2013). Knowledge of the host location mechanism employed by this species would be useful for developing methods for applying them as biological control agents.

Abundant information is available on biological characteristic, predatory behavior, and artificial rearing of predaceous stink bugs (Streams *et*

al. 1963, Zahn *et al.* 2008, De Bortoli *et al.* 2011, Helmey-Hartman & Miller 2014). Mechanisms underlining mate location and recognition are still unknown for most stink bugs. Research on stink bug mating behavior has indicated the importance of antennae in mate location and recognition. Numerous studies have characterized antennal sensilla of various insects (Zacharuk 1980, Keil & Steinbrecht 1984, Städler 1984, Cõnsoli *et al.* 1999, Bleeker *et al.* 2004, Bourdais *et al.* 2006, Yu *et al.* 2013) whereas very few of such studies have focused on Pentatomidae (Rani & Madhavendra 1995, Brézot *et al.* 1997, Silva *et al.* 2010).

Insect antennae play important roles in various behaviors, including habitat searching, host location, discrimination, courtship and mating behavior (Schneider 1964). Antennae of insects contain sensory sensilla, which function in the detection of various stimuli involved in the host habitat and mate location (Chapman 1998). As part of our ongoing research on mate and host location mechanisms of *A. chinensis*, we characterized and determined the abundance and distribution of the antennal sensilla of this stink bug using the techniques of scanning electron microscopy (SEM). This work is a prerequisite for future electrophysiological studies of the antennal sensory system involved in intraspecific chemical communication. The types of sensilla of *A. chinensis* are also compared with those described for other heteropteran insects (e.g., Rani & Madhavendra 1995, Brézot *et al.* 1997, Chinta *et al.* 1997, Rani & Madhavendra 2005, Silva *et al.* 2010).

2. Material and methods

The morphology and distribution of sensilla on the antennae of *A. chinensis* were investigated. Sensilla on the dorsal and ventral surfaces of the antennae of both sexes were identified, counted, and measured. To characterize the sensilla we used the nomenclature proposed by Schneider (1964) and Zacharuk (1980, 1985). The following abbreviations are used for different types of sensilla:

ST1: Sensilla trichoidea 1, ST2: Sensilla trichoidea 2, SB1: Sensilla basiconica 1, SB2:

Sensilla basiconica 2, SB3: Sensilla basiconica 3, SB4: Sensilla basiconica 4, SCH: Sensilla chaetica, SCA: Sensilla cavity, SCO: Sensilla coelomic.

2.1. Insects

Adults of *A. chinensis* were initially collected from *Ulmus macrocarpa* Hance trees, located in Qian'an County, Jilin Province, northeastern China, and subsequently maintained in the laboratory for several generations. *Arma chinensis* were fed with Chinese oak silk moth, *Antheraea pernyi*, pupae and reared at $27 \pm 1^\circ\text{C}$, RH of $75 \pm 5\%$, and a 16:8 (L:D) hour photoperiod. The insects were provided regularly with water. Only adult insects, 1 week old or older, were used in the experiments. The heads of the test insects were excised from the live insects and prepared for examination under the scanning electron microscope. Voucher specimens were deposited in Institute of Forest Protection, Jilin Provincial Academy of Forestry, Changchun, China.

2.2. Scanning electron microscopy (SEM)

Antennae of specimens were carefully excised from the antennal sockets with fine forceps under a stereomicroscope (Olympus SZX12, Japan). The antennae were first kept in 70% ethanol for 24h and then dehydrated in a graded alcohol series of 75, 80, 85, 90, and 100% in each case for 10 min each. The antennae were individually mounted with dorsal or ventral sides on aluminum stubs with double-sided sticky tapes. Before examination the antennae were sputter-coated with gold (20 nm) in a Hitachi E-102 high resolution sputter coater. The specimens were examined in a Hitachi S-570 scanning electron microscope (Hitachi, Tokyo, Japan) set at 20kV. 10 antennae of each sex were examined under the SEM.

2.3. Statistical analysis

The data were analyzed using the t-test with the statistical program SPSS version 17.0 (SPSS Inc., Chicago, IL) for Windows.

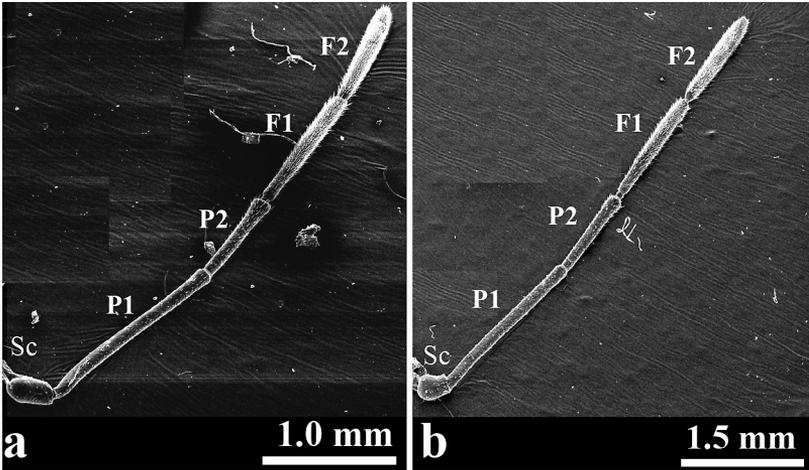


Fig 1. General view of antenna of *Arma chinensis* showing scape (Sc), segments of pedicel (P1–2) and flagellum (F1–2). – a. Male. – b. Female.

Table 1. Means (\pm SD) of lengths (mm) of antennal segments of *Arma chinensis* ($n=10$ per sex).

Sex	Scape	Pedicel 1	Pedicel 2	Flagellum 1	Flagellum 2	Total*
Female	0.402 \pm 0.015	1.772 \pm 0.062	0.730 \pm 0.045	1.158 \pm 0.077	0.946 \pm 0.034	5.008 \pm 0.233
Male	0.368 \pm 0.031	1.535 \pm 0.031	0.772 \pm 0.034	1.163 \pm 0.025	1.055 \pm 0.032	4.893 \pm 0.153

* Significant difference between the sexes (Student's t-test, $p<0.05$).

Table 2. Means (\pm SD) of lengths and diameters (μ m) as well as morphological characteristics of different types of antennal sensilla in *A. chinensis* ($n=10$ per sex).

Type*	Sex	Length**	Diameter**	Morphological characteristics			
				Tip	Wall	Shape	Socket
ST1	Female	44.60 \pm 1.75	3.31 \pm 0.22	Blunt	Smooth	Straight or curved	Wide
	Male	45.46 \pm 2.73	3.02 \pm 0.05				
ST2	Female	38.70 \pm 1.25	1.91 \pm 0.07	Blunt	Smooth	Straight or curved	Wide
	Male	40.66 \pm 2.77	2.13 \pm 0.11				
SB1	Female	16.96 \pm 2.64	2.55 \pm 0.22	Blunt	Smooth	Straight	Tight
	Male	15.45 \pm 4.33	2.28 \pm 0.48				
SB2	Female	35.48 \pm 11.37	3.35 \pm 0.47	Blunt	Smooth	Straight	Tight
	Male	37.23 \pm 4.77	3.88 \pm 0.53				
SB3	Female	13.17 \pm 3.83	2.29 \pm 0.47	Blunt	Grooved	Curved	Tight
	Male	11.3 \pm 2.55	2.14 \pm 0.33				
SB4	Female	6.87 \pm 2.02	1.73 \pm 0.34	Blunt	Smooth	Straight	Tight
	Male	8.41 \pm 1.12	1.69 \pm 0.21				
SCH	Female	55.49 \pm 12.01	4.58 \pm 0.43	Blunt	Grooved	Straight or curved	Wide
	Male	43.66 \pm 14.86	4.25 \pm 0.90				
SCA	Female	–	1.61 \pm 0.21	–	–	–	–
	Male	–	1.55 \pm 0.30				
SCO	Female	1.20 \pm 0.07	2.38 \pm 0.23	–	–	–	–
	Male	1.16 \pm 0.15	2.35 \pm 0.20				

* Abbreviations: ST1: Sensilla trichoidea 1; ST2: Sensilla trichoidea 2; SB1: Sensilla basiconica 1; SB2: Sensilla basiconica 2; SB3: Sensilla basiconica 3; SB4: Sensilla basiconica 4; SCH: Sensilla chaetica; SCA: Sensilla cavity; SCO: Sensilla coeloconic.
 ** No significant differences between the sexes (Student's t-test, all $p>0.05$).

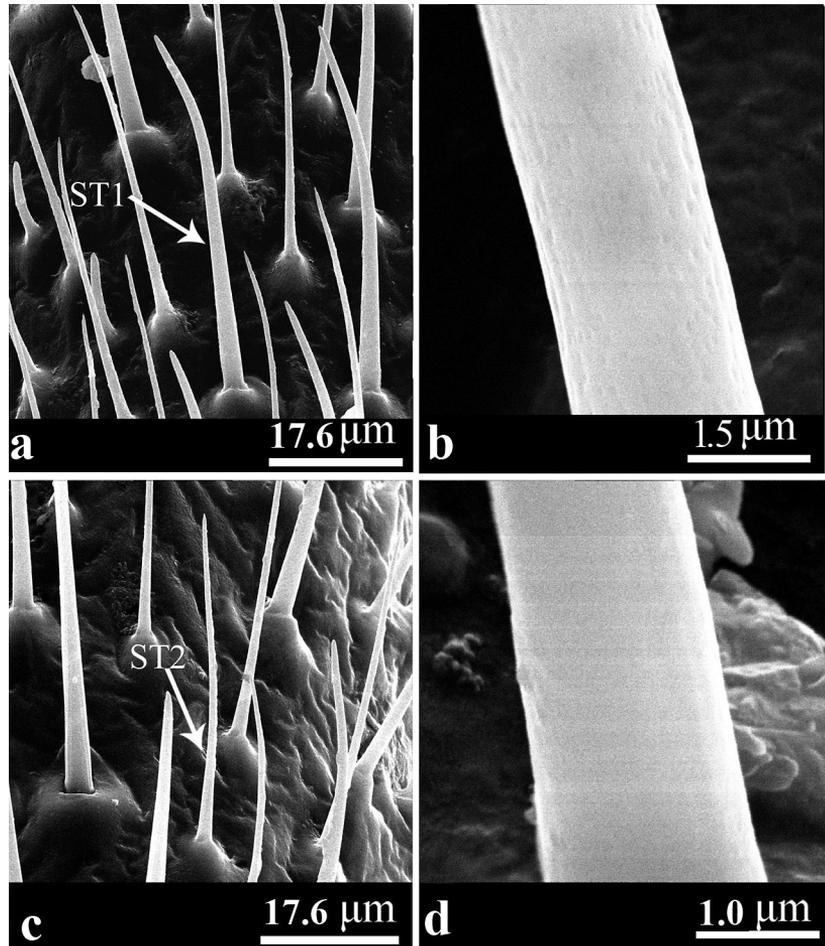


Fig 2. Details of antenna of *A. chinensis*. – a. ST1 (sensilla trichodea 1) on the ventral side of the 1st flagellomere of male. – b. The smooth cuticle of ST1 at higher magnification. – c. ST2 (sensilla trichodea 2) on the ventral side of the 1st flagellomere of male. – d. The smooth cuticle of ST2 at higher magnification.

3. Results

3.1. General structure of the antennae

Antennae of both sexes of *A. chinensis* were morphologically similar. The filiform antennae were comprised of a basal scape (Sc), a two-segmented pedicel (P1, P2) and a two-segmented flagellum (F1, F2) (Fig. 1). Female adults bear much longer antennae than the males. The mean lengths of the antennal segments of *A. chinensis* are summarized in Table 1.

3.2. Types of sensilla

Based on their morphology, two types of sensilla trichodea (ST1-2), four types of sensilla basiconica (SB1-4), one type of sensilla chaetica (SCH), one type of sensilla cavity (SCA) and one

type of sensilla coeloconica (SCO) were distinguished on the antennae in both sexes. The characteristic morphological features of the antennal sensilla of *A. chinensis* are summarized in Table 2. The type and size of the sensilla covering the different parts of the antenna were similar in the female and male stink bugs, and no sexual dimorphism was found in these sensitive structures (Table 2). The counts of sensilla were made from both the dorsal and ventral sides.

3.2.1. Sensilla trichodea 1 (ST1)

ST1 were long, straight or curved hairs, with their bases tightly inserted in an elevation of the cuticle, with a pointed tip (Fig. 2a). The sensilla stood up perpendicularly to the antennal surface and when viewed at a high magnification their wall was smooth with no pores or grooves on the sur-

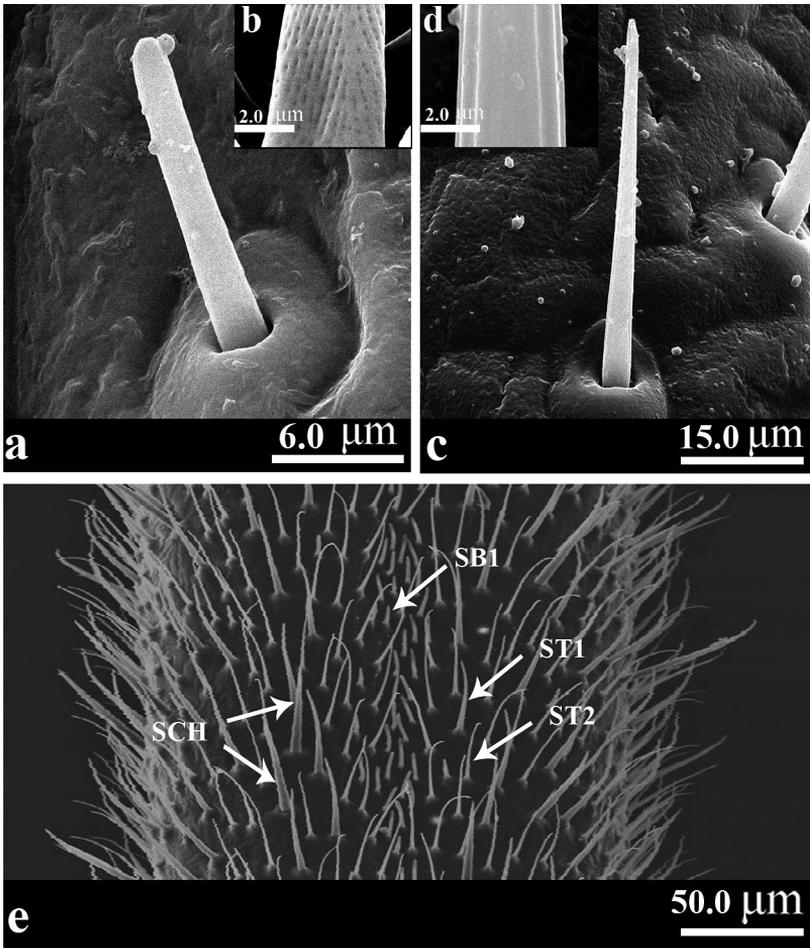


Fig 3. Details of antenna of *A. chinensis*. – a. SB1 (sensilla basiconica 1) on the dorsal side of the 1st pedicel of male. – b. Numerous pores on the cuticle of SB1 at higher magnification. – c. SB2 (sensilla basiconica 2) on the ventral side of the 1st flagellomere of female. – d. Longitudinal grooves on the wall of SB2. – e. SB1 on the central axis of the dorsal side of the 2nd flagellomere of female.

face (Fig. 2b). They were found scattered on the flagellum of the antennae. ST1 was absent from the scape and the pedicel. This type of sensillum had a mean length and basal diameter (\pm SD) of $44.60 \pm 1.75 \mu\text{m}$ and $3.31 \pm 0.22 \mu\text{m}$ (female) and $45.46 \pm 2.73 \mu\text{m}$ $3.02 \pm 0.05 \mu\text{m}$ (male), respectively.

3.2.2. *Sensilla trichodea 2 (ST2)*

ST2 were long and slender hairs with straight or slightly curved shafts, and tapering to a sharp point apically (Fig. 2c). The shaft of this sensillum was smooth, arose from an elevated base and did not show any pores (Fig. 2d). They were located only on the flagellum of the antennae. ST2 were on average (\pm SD) $38.70 \pm 1.25 \mu\text{m}$ (female) and $40.66 \pm 2.77 \mu\text{m}$ (male) in length and $1.91 \pm 0.07 \mu\text{m}$ (female) and $2.13 \pm 0.11 \mu\text{m}$ (male) in basal diameter.

3.2.3. *Sensilla basiconica 1 (SB1)*

SB1 were stout pegs with slightly curved shaft having numerous pores on the walls (Figs. 3a, b). These blunt-tipped sensilla inserted into sockets often formed by slightly elevated craters. These sensilla were found on the central axis of the surface of the terminal antennal segments (Fig. 3e). SB1 measured on average (\pm SD) $16.96 \pm 2.64 \mu\text{m}$ (female) and $15.45 \pm 4.33 \mu\text{m}$ (male) in length, $2.55 \pm 0.22 \mu\text{m}$ (female) and $2.28 \pm 0.48 \mu\text{m}$ (male) in basal diameter.

3.2.4. *Sensilla basiconica 2 (SB2)*

SB2 had blunt tips and a well defined socket at the base. They were shorter and thicker at the base than SB1 (Fig. 3c). They were present along the entire length of the antennae of both sexes. But

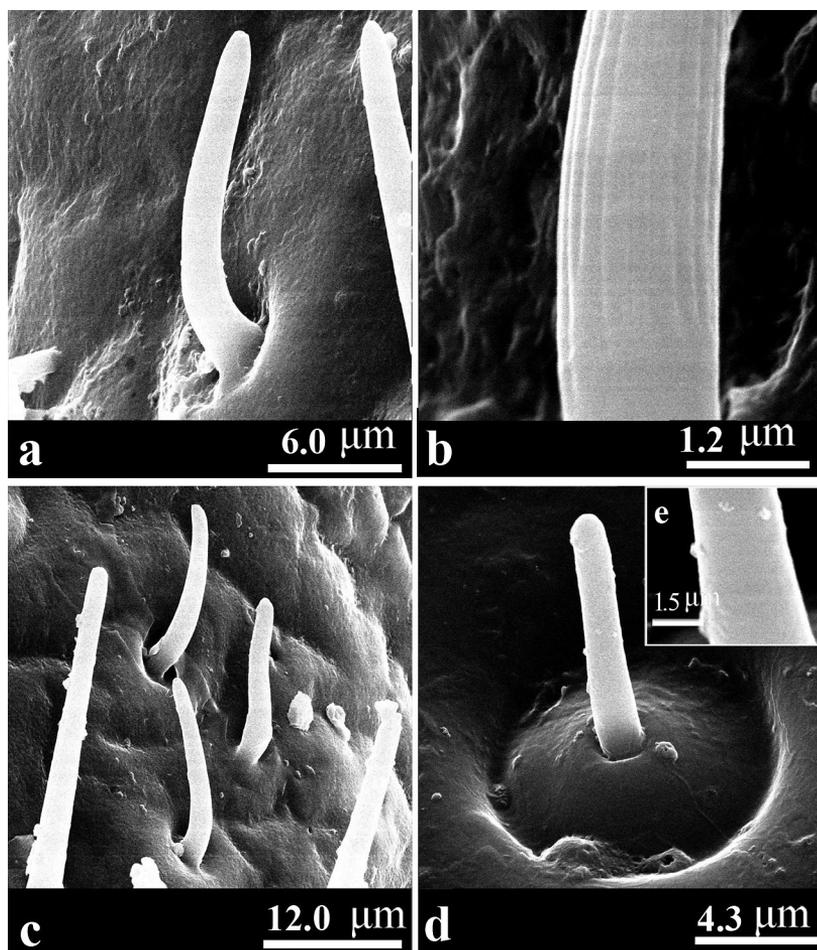


Fig 4. Details of antenna of *A. chinensis*. – a. SB3 (Sensilla basiconica 3) on the ventral side of the 2nd pedicel of male. – b. Longitudinal grooves on the wall of SB3. – c. A dense group of SB3 on the 2nd pedicel of male. – d. SB4 (sensilla basiconica 4) on the dorsal side of the scape of male. – e. The smooth cuticle of SB4 at higher magnification.

analyzing the SB2 at high magnification we observed some longitudinal grooves on the cuticle (Fig. 3d). They measured on average (\pm SD) $35.48 \pm 11.37 \mu\text{m}$ (female) and $37.23 \pm 4.77 \mu\text{m}$ (male) in length and 3.35 ± 0.47 (female) and $3.88 \pm 0.53 \mu\text{m}$ (male) in basal diameter.

3.2.5. *Sensilla basiconica 3 (SB3)*

SB3 abruptly curved in the direction of the antennal surface and were on average (\pm SD) $13.17 \pm 3.83 \mu\text{m}$ (female) and $11.3 \pm 2.55 \mu\text{m}$ (male) long pegs with a smooth wall and blunt tip (Figs. 4a, b). Their diameter at the base was $2.29 \pm 0.47 \mu\text{m}$ (female) and $2.14 \pm 0.33 \mu\text{m}$ (male). They were usually situated as a dense group (Fig. 4c) of 3–7 sensilla and there were usually $8.34 \pm 3.20 \mu\text{m}$ between separate sensilla.

3.2.6. *Sensilla basiconica 4 (SB4)*

The mean (\pm SD) lengths of SB4 were $6.87 \pm 2.02 \mu\text{m}$ (female) and $8.41 \pm 1.12 \mu\text{m}$ (male). They were blunt-tipped pegs with basal diameters of $1.73 \pm 0.34 \mu\text{m}$ (female) and $1.69 \pm 0.21 \mu\text{m}$ (male) and possessed no grooves or pores on the antennal surface (Fig. 4d, e). They were typically inserted into a small dome, bore no articulating socket, and were oriented perpendicularly to the antennal surface. They were situated on the surface of the scape, pedicel, and the first flagellomere.

3.2.7. *Sensilla chaetica (SCH)*

SCH were long sickle-shaped strong bristles with longitudinal grooves accumulating toward the tip, which were located in an open articulating

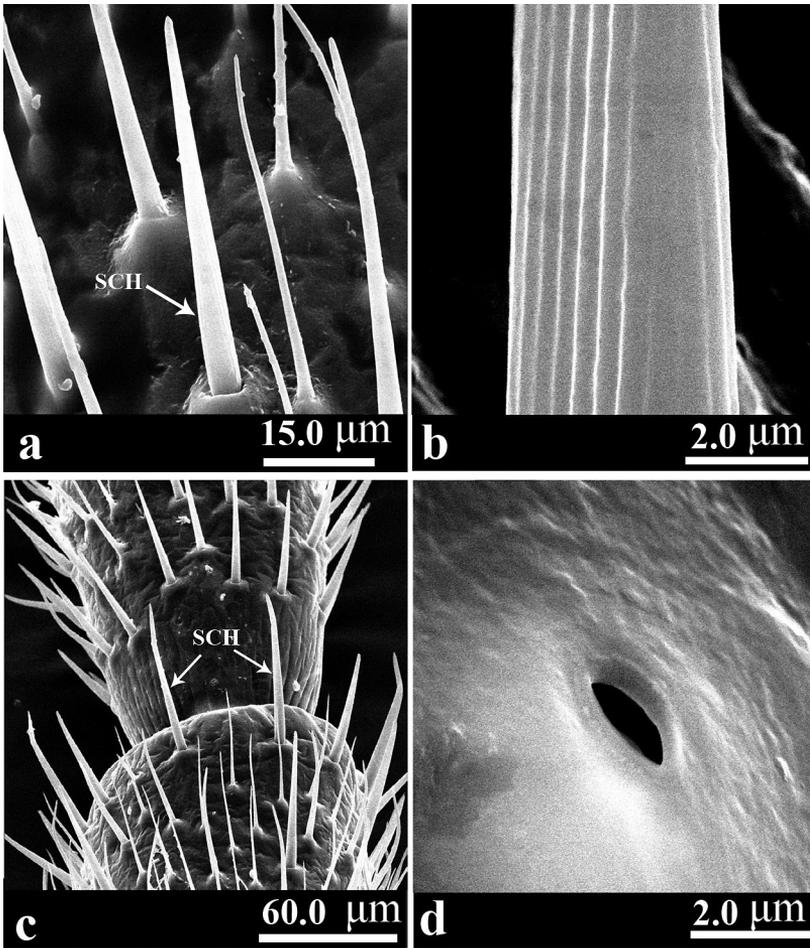


Fig 5. Details of antenna of *A. chinensis*. – a. SCH (sensilla chaetica) on the ventral side of the 2nd flagellomere of female. – b. Longitudinal grooves on the wall of SCH. – c. SCH located on the distal area of the dorsal side of the 1st flagellomere. – d. SCA (Sensilla cavity) on the dorsal side of the 2nd flagellomere of male.

socket (Figs. 5a, b). These sensilla lay parallel to the surface and point towards the tip of the antenna. They covered the second pedicel and the two flagellomeres (Fig. 5c). On average (\pm SD), SCH measured 55.49 ± 12.01 μm (female) and 43.66 ± 14.86 μm (male) in length and 4.58 ± 0.43 μm (female) and 4.25 ± 0.90 μm (male) in basal diameter.

3.2.8. Sensilla cavity (SCA)

SCA were single large pores that occurred at the base of the second flagellomere (Fig. 5d). They were on average (\pm SD) 1.61 ± 0.21 μm (female) and 1.55 ± 0.30 μm (male) in the diameter. They were situated not only on the dorsal, but also on the ventral side of the second flagellomere. Due to their small numbers these sensilla were not considered in the sensilla counts.

3.2.9. Sensilla coeloconic (SCO)

SCO were short peg-like sensory structures sunk into a pit with the apex of the peg not projecting out of the pit (Fig. 6a). They occurred exclusively on the second segment of the flagellum and had a mean (\pm SD) diameter of 2.38 ± 0.23 μm (female) and 2.35 ± 0.20 μm (male). These sensilla were not considered in the sensilla counts because of their limited number.

3.3. Abundance and distribution of antennal sensilla

The mean numbers and distribution of sensilla on the segments of *A. chinensis* are recorded in Table 3. Sensilla were absent on the segmental joint between the second pedicel and the first flagellomere-

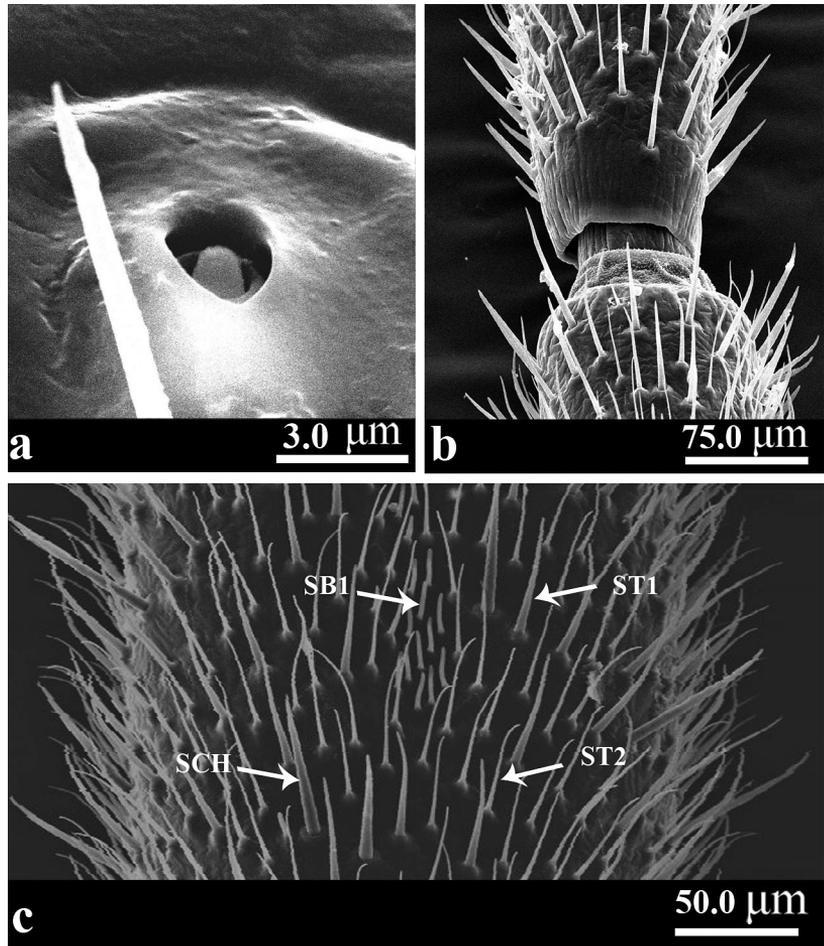


Fig 6. Details of antenna of *A. chinensis*. – a. SCO (sensilla coeloconica) on the dorsal side of the 2nd flagellomere of male. – b. Segmental joint of the 2nd pedicel and the 1st flagellomere in male, showing absence of sensilla. – c. Large amount of different types of sensilla on the dorsal side of the 2nd flagellomere of female: SB1: Sensilla basiconica 1, ST1: Sensilla trichoidea 1, SCH: Sensilla chaetica, ST2: Sensilla trichoidea 2.

re of both sexes (Fig. 6b). Large amounts of different types of sensilla were distributed on the dorsal and ventral side of the flagellum (Fig. 6c). The distribution patterns of sensilla on the antennae of both sexes of the stink bug were similar. ST1 and ST2 were more abundant than the other types of sensilla on the antennae. SB1 were absent from the scape. SB2 were distributed throughout the antennae. SB3 were distributed on the second segment of the pedicel and the two segments of the flagellum. SB4 were absent from the second flagellum. ST1 and ST2 were absent from the scape and pedicel. SC were located on the second segment of the pedicel and the two segments of the flagellum. SCA and SCO only occurred on the second segment of the flagellum.

4. Discussion

The various types of sensilla and their distribution on the antennae of male and female *A. chinensis* as revealed in this study were similar with those reported for other heteropteran insect species (Rani & Madhavendra 1995, Catalá, 1997, Onagbola *et al.* 2008). The antennae of insects have been typically described as consisting of the three usual components: scape, pedicel, and flagellum (Chapman 1998). The flagellum of *A. chinensis* is formed of two segments that hold the largest number of sensilla among the different segments of the antenna. Similarly, two segments were observed in *Neomegalotomus parvus* (Westwood) (Ventura & Panizzi 2005) (Alydidae), *Panstrongylus megistus* (Burmeister) (Vilella *et al.* 2005) and *Rhodnius prolixus* (Stal) (Akent'eva 2008) (Reduviidae), *Coreus margi-*

Table 3. Mean numbers per segment (\pm SD) and distribution of different types of antennal sensilla of female and male *A. chinensis* ($n=5$ antennae per sex). For abbreviations, see Table 2.

Type	Sex	Scape	Pedicel 1	Pedicel 2	Flagellum 1	Flagellum 2	Total
ST1 ^{ns}	Female	0	0	0	242.0 \pm 21.5	270.0 \pm 19.5	542.3 \pm 44.0
	Male	0	0	0	193.3 \pm 10.5	420.0 \pm 30.4	613.3 \pm 40.9
ST2 ^{ns}	Female	0	0	0	493.6 \pm 20.5	521.6 \pm 19.8	1015.3 \pm 40.3
	Male	0	0	0	317.6 \pm 21.1	611.3 \pm 25.4	928.9 \pm 46.5
SB1 ^{ns}	Female	0	110.6 \pm 11.1	63.3 \pm 6.0	20.6 \pm 2.5	127.6 \pm 12.5	322.3 \pm 32.1
	Male	0	114.3 \pm 11.0	26.0 \pm 6.0	58.6 \pm 17.0	148.6 \pm 12.1	347.6 \pm 46.1
SB2 ^{ns}	Female	6.0 \pm 0	65.0 \pm 8.0	96.6 \pm 8.7	306.3 \pm 19.2	124.3 \pm 5.1	598.3 \pm 41.1
	Male	5.0 \pm 0	41.0 \pm 7.2	99.0 \pm 5.5	186.6 \pm 10.7	144.0 \pm 5.5	475.6 \pm 29.1
SB3*	Female	0	0	7.0 \pm 2.0	26.6 \pm 1.5	8.3 \pm 1.5	41.9 \pm 5.0
	Male	0	0	8.3 \pm 2.5	19.6 \pm 1.5	45.0 \pm 11.1	72.9 \pm 15.1
SB4 ^{ns}	Female	30.3 \pm 3.0	81.0 \pm 4.0	23.6 \pm 2.1	15.6 \pm 3.1	0	120.3 \pm 9.1
	Male	31.6 \pm 3.1	59.3 \pm 13.2	6.0 \pm 0	35.6 \pm 4.0	0	132.6 \pm 20.2
SCH*	Female	0	0	16.0 \pm 3.0	49.3 \pm 8.1	75.6 \pm 13.0	140.9 \pm 24.0
	Male	0	0	12.6 \pm 2.5	108.3 \pm 11.0	92.6 \pm 6.5	213.6 \pm 20.0

* Significant difference in total number of antennal sensilla between the sexes (Student's t-test, $p < 0.05$).

ns = No significant difference in total number of antennal sensilla between the sexes (Student's t-test, $p > 0.05$).

natus (Linnaeus) (Aken't'eva 2008) and *Leptoglossus zonatus* (Dallas) (Gonzaga-Segura *et al.* 2013) (Coreidae), and *Nezara viridula* (Linnaeus) (Brézot *et al.* 1997) (Pentatomidae). Gonzaga-Segura *et al.* (2013) inferred that this could be an adaptation to improve olfaction. Our study revealed nine morphologically different types of sensilla on the antennae of both sexes of *A. chinensis*, similar to those described for other stink bugs (Brézot *et al.* 1997, Chinta *et al.* 1997, Rani & Madhavendra 2005, Silva *et al.* 2010). The presence of sensilla campaniform and sensilla placoid found on other heteropteran insects (Gonzaga-Segura *et al.* 2013) were not found on *A. chinensis*.

Sensilla trichodea 1 (ST1) resemble the “type 5 sensilla” in the green stink bug, *N. viridula* (Brézot *et al.* 1997); “long straight hair type 4” in the tarnished plant bug, *Lygus lineolaris* (Palisot de Beauvois) (Chinta *et al.* 1997); “thick-walled trichoid sensilla” in the *P. megistus* (Catalá, 1997); “aporous sensilla trichodea type II” in the Asian citrus psyllid, *Diaphorina citri* Kuwayama (Onagbola *et al.* 2008); “trichoid sensilla 1” in *Euschistus heros* (Fabricius), *Piezodorus guildinii* (Westwood) and *Edessa meditabunda* (Fabricius) (Silva *et al.* 2010). Sensilla trichodea 2 (ST2) with blunt tip in *A. chinensis* appear to be identical with the “sensilla trichodea type I” in *Odontopus nigricornis* (Stall) (Rani & Madha-

vendra 1995); “type I sensilla” in the green stink bug, *N. viridula* (Brézot *et al.* 1997) and “thin-walled trichoid sensilla” in the *P. megistus* (Catalá 1997). They are the most abundant type of sensilla present on the antennae of these and many other stink bugs. Shorter or longer blunt-tipped hairs, which do not articulate at their base, like the sensilla trichodea (ST) in *A. chinensis*, occur abundantly on the antennae of many wasps, beetles and moths (Bleeker *et al.* 2004, Merivee *et al.* 1999, Lopes *et al.* 2005, Castrejón Gómez & Carrasco 2008). Sensilla trichodea have traditionally been thought of as the contact chemoreceptors (Schneider 1964). In *N. viridula*, they have been reported to be good candidates for contact chemoreception based on their external structure (Brézot *et al.* 1997). Bioassay experiments of the mating behavior on *Phoracantha semipunctata* (Fabricius) demonstrated that sensilla trichodea served as a contact chemoreceptor (Lopes *et al.* 2005). Because accounting for more than 50% of the total number of the antennal sensilla, sensilla trichodea can be considered the main sensory receptors involved in the chemical sense in neotropical bugs (Silva *et al.* 2010). ST was the most abundant of all sensillum types on the antennae of both sexes of *A. chinensis*, and they were also thought to function as chemoreceptors (Zacharuk 1985).

Large articulated bristles with blunt tip, SCH

in *A. chinensis*, seem to be identical with the "bristle-like type 3" in *N. viridula* (Brézot *et al.* 1997), "long curved bristle type 2" in *L. lineolaris* (Chinta *et al.* 1997). Brézot *et al.* (1997) suggested that the flexible socketed base, the length of the hair, the dendrite not protruding in the hair, the presence of a cuticular shaft under the cuticle near the base of this type of sensillum are characteristics generally correlated with mechanoreception in *N. viridula*. In *L. lineolaris*, the presence of a thick cuticle and a single terminal pore of this type of sensillum indicated its possible involvement in chemoreception in contacts (Chinta *et al.* 1997). In *Monochamus notatus* (Drury) and *M. scutellatus* (Say), they can be considered as the receptors responding to sound, wind, or touch (Dyer & Seabrook 1975). The absence of pores in this sensilla precludes a chemosensory role in *A. chinensis* (Fig. 5b). SCH lacked pores and protruded above all other sensilla, so we assume that they function as mechanoreceptor in *A. chinensis*.

Sensilla basiconica 1 (SB1) with multiple pores and without flexible sockets of *A. chinensis* look like the "sensilla basiconica type I" in the stink bug *O. nigricornis* (Rani & Madhavendra 1995); "type 4 sensilla" in the green stink bug, *N. viridula* (Brézot *et al.* 1997); "short peg" in the tarnished plant bug, *L. lineolaris* (Chinta *et al.* 1997); "sensilla basiconica type 1" in the ground beetle, *Bembidion properans* Stephens (Merivee *et al.* 2002); "basiconic sensilla type 3" in *E. heros*, *P. guildinii* and *E. meditabunda* (Silva *et al.* 2010). SB1 should correspond to the "thin-walled" (Slifer 1970), "single-walled, wall pore sensilla" (Altner & Prillinger 1980) and "multiporous chemosensilla" (Zacharuk 1980). Sensilla basiconica bear structural features such as a non-flexible base, a thin, multiporous cuticular wall, a pore-tubule system and branched dendritic segments (Zacharuk 1980). The numerous pores and branched dendrites are traditionally considered to provide evidence that these sensilla basiconica function as olfactory receptors (Altner & Prillinger 1980, Zacharuk 1985). The cuticular wall of SB1 bears numerous pores in *A. chinensis* (Fig. 2b). Based on the ultrastructure observed by SEM, SB1s are inferred as the olfactory receptors in *A. chinensis*. Sexual dimorphism in the numbers of these sensilla was observed in *L. lineolaris* (Chinta *et al.* 1997). In the species studied here

the number of SB1 was approximately equal between the males and the females (Table 3). In other insects, these basiconic sensilla are known to be olfactory receptors responding to food and habitat related compounds (Schneider 1964, Zacharuk 1985).

The clusters of SB1 are located on the distal part of the second flagellomere (Fig. 3e). Clusters of olfactory sensilla basiconica have been identified also in many coleopteran species, such as *M. notatus* and *M. scutellatus* (Dyer & Seabrook 1975), *Hypera meles* (Fabricius) (Smith *et al.* 1976), *Psacotheta hilaris* (Pascoe) (Dai & Honda 1990), *Phyllotera cruciferae* (Goeze), *Psylliodes punctulata* Melsheimer, *Epitrix cucumeris* (Harris) and *Psylliodes affinis* (Paykull) (Ritcey & McIver 1990), and *Geotrupes auratus* Motschulsky and *Copris pecuarius* Lewis (Inouchi *et al.* 1987). It has been inferred that such olfactory sensilla clusters function as sensory fields (Dyer & Seabrook 1975) and may compose an enlarged odor-sensing area that would be advantageous for long-distance olfactory detection (Inouchi *et al.* 1987).

Sensilla basiconica 2 (SB2) of *A. chinensis* are common on the antennae of many stink bugs. These sensilla resemble the "type 2 sensilla" of *N. viridula* (Brézot *et al.* 1997), "basiconic sensilla" of *Riptortus pedestris* Fabricius, *Elasmolomus sordidus* (Fabricius), *Cyclopelta siccifolia* Westwood and *Chrysocoris purpurea* (Westwood) (Rani & Madhavendra 2005), and "basiconic sensilla 2" of *E. heros*, *P. guildinii* and *E. meditabunda* (Silva *et al.* 2010). Silva *et al.* (2010) suggested that this type of sensillum may be related to finding food or suitable habitats. Some authors have assigned a thermo- or chemoreception function to these sensilla (Chapman 1982, Zacharuk 1985).

Sensilla basiconica 3 (SB3) of *A. chinensis* are very similar to the "sensilla basiconica type 1" of the ground beetle *Platynus dorsalis* (Pontopidan) (Merivee *et al.* 2001) and "sensilla basiconica type 2" in the ground beetle, *B. properans* (Merivee *et al.* 2002). Non-articulated blunt-tipped basiconic sensilla are common on the antennal flagellum of most insects. In the ground beetle *Nebria brevicollis* (Fabricius) and the cigarette beetle *Lasioderma serricorne* (Fabricius), it was demonstrated that the wall of

these sensilla is perforated by numerous tiny pores (Daly & Ryan 1979, Okada *et al.* 1992). The numerous pores and branched dendrites are considered to be evidence that these basiconic sensilla function as olfactory receptors (Altner & Prillinger 1980, Zacharuk 1985). In this study, we did not detect pores in the cuticle of SB3 (Fig. 4b). Single cell recording studies and behavioral experiments are needed to confirm their function.

The appearance and small number of sensilla basiconica 4 (SB4) of *A. chinensis* are similar with “sensilla basiconica type VII” in the click beetle, *Agriotes obscurus* Linnaeus (Merivee 1992), “sensillum styloconicum” in the cigarette beetle *L. serricornis* (Okada *et al.* 1992), “double-walled sensilla” in *Ips typographus* (Linnaeus) (Hallberg 1982), “sensilla basiconica II” in the ladybird beetle *Semiadalia undecimnotata* Schneider (Jourdan *et al.* 1995). Sensilla without pores are either mechanosensitive or thermo/hygro-sensitive in *A. chinensis* (Altner & Prillinger 1980, Keil 1999).

Sensilla cavity (SCA) in *A. chinensis* are similar to those of “cavity sensillum” of eight Acrididae species (Li *et al.* 2007), “sensilla coeloconica” of *B. properans* (Merivee *et al.* 2002), “sensilla cavity” of *Callosobruchus chinensis* (Linnaeus) and *C. maculatus* (Fabricius) (Hu *et al.* 2009), “sensilla cavity” of *Cnaphalocrocis medinalis* (Guenée) (Sun *et al.* 2011) and probably in other heteropteran species as well. Chemo-, thermo-, or hygroreception is the most probable function for these sensilla (Zacharuk 1985). They were inferred to function as contact-chemoreceptors, allowing the insect to obtain information on the suitability of any plant they touch (Sun *et al.* 2011).

Sensilla coeloconica (SCO) were found only on the second segment of the flagellum of *A. chinensis*. Sensilla with similar morphological features were found on the flagellum of *N. viridula*, *C. purpurea* (West), *E. heros*, *P. guildinii* and *L. zonatus* (Brézot *et al.* 1997, Rani & Madhavendra 2005, Silva *et al.* 2010, Gonzaga-Segura *et al.* 2013) and Reduviidae (Catalá 1997). In social insects, such as ants and wasps, whose behavior is guided by variations in temperature, sensilla coeloconica are chemoreceptors that respond to air temperature changes (Ruchty *et al.* 2009). In lepidopterans these sensilla are olfactory receptors,

possibly sensitive to volatile plant odors (Van der Pers 1981, Pophof 1997), and in homopterans they function as hygroreceptors preventing desiccation of the antennae (Kristoffersen *et al.* 2006).

Further studies on functional antennal morphology and anatomy are needed to confirm the proposed functions of the sensilla identified in this study. The present results provide necessary background information for our ongoing study on electrophysiology and chemical ecology of *A. chinensis*.

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References

- Akent'eva, N. A. 2008: The formation of the antenna sensory apparatus in some bug (Heteroptera) species in the course of their postembryonic development. — *Entomological Review* 88: 381–390.
- Altner, H. & Prillinger, L. 1980: Ultrastructure of invertebrate chemo-, thermo-, and hygroreceptors and its functional significance. — *International Review of Cytology* 67: 69–139.
- Bleeker, M. A. K., Smid, H. M., van Aels, A. C., van Loon J. J. A. & Vet L. M. E. 2004: Antennal sensilla of two parasitoid wasps: A comparative scanning electron microscopy study. — *Microscopy Research and Technique* 63: 266–273.
- Bourdais, D., Vernon, P., Krespi, L., Lannic, J. L. & Van Baaren, J. 2006: Antennal structure of male and female *Aphidius rhopalosiphii* DeStefani-Peres (Hymenoptera: Braconidae): Description and morphological alterations after cold storage or heat exposure. — *Microscopy Research and Technique* 69: 1005–1013.
- Brézot, P., Tauban, D. & Renou, M. 1997: Sense organs on the antennal flagellum of the green stink bug, *Nezara viridula* (L.) (Heteroptera: Pentatomidae): sensillum types and numerical growth during the post-embryonic development. — *International Journal of Insect Morphology and Embryology* 25: 427–441.
- Castrejón Gómez, V. R. & Carrasco, J. V. 2008: Morphological characteristics of Antennal Sensilla in *Talponia batesi* (Lepidoptera: Tortricidae). — *Annals of the Entomological Society of America* 101: 181–188.
- Catalá, S. S. 1997: Antennal sensilla of Triatominae (Hemiptera: Reduviidae): a comparative study of five gen-

- era. — *International Journal of Insect Morphology and Embryology* 26: 67–73.
- Chapman, R. F. 1982: Chemoreception: The significance of receptor numbers. — *Advances in Insect Physiology* 16: 247–356.
- Chapman, R. F. 1998: *The insects: Structure and function*. 4th Edition. — Cambridge University Press, New York. 788 pp.
- Chinta, S., Dickens, J. C. & Baker, G. T. 1997: Morphology and distribution of antennal sensilla of the tarnished plant bug, *Lygus lineolaris* (Palisot de Beauvois) (Hemiptera: Miridae). — *International Journal of Insect Morphology and Embryology* 26: 21–26.
- Cónsoli, F. L., Kitajima, E. W. & Postalí Parra, J. R. 1999: Sensilla on the antenna and ovipositor of the parasitic wasps *Trichogramma galloi* Zucchi and *T. pretiosum* Riley (Hymenoptera: Trichogrammatidae). — *Microscopy Research and Technique* 45: 313–324.
- Dai, H. & Honda, H. 1990: Sensilla on the antennal flagellum of the yellow spotted longicorn beetle, *Psacotheta hilaris* (Pascoe) (Coleoptera: Cerambycidae). — *Applied Entomology and Zoology* 25: 273–295.
- Daly, P. J. & Ryan, M. F. 1979: Ultrastructure of antennal sensilla of *Nebria brevicollis* (Fab.) (Coleoptera: Carabidae). — *International Journal of Insect Morphology and Embryology* 8: 169–181.
- De Bortoli, S. A., Otuka, A. K., Vacari, A. M., Martins, M. I. E. G. & Volpe, H. X. L. 2011: Comparative biology and production costs of *Podisus nigrispinus* (Hemiptera: Pentatomidae) when fed different types of prey. — *Biological Control* 58: 165–174.
- Dyer, L. J. & Seabrook, W. D. 1975: Sensilla on the antennal flagellum of the Sawyer Beetles *Monochamus notatus* (Drury) and *Monochamus scutellatus* (Say) (Coleoptera: Cerambycidae). — *Journal of Morphology* 146: 513–532.
- Gonzaga-Segura, J., Carrasco, J. V. & Castrejón Gómez, V. R. 2013: Sense Organs on the Antennal Flagellum of *Leptoglossus zonatus* (Heteroptera: Coreidae). — *Annals of the Entomological Society of America* 106: 511–517.
- Hallberg, E. 1982: Sensory organs in *Ips typographus* (Insecta: Coleoptera) fine structure of antennal sensilla. — *Protoplasma* 111: 206–214.
- Helmey-Hartman, W. L. & Miller, C. W. 2014: Context-Dependent Mating Success in *Murgantia histrionica* (Hemiptera: Pentatomidae). — *Annals of the Entomological Society of America* 107: 264–273.
- Hu, F., Zhang, G. N. & Wang, J. J. 2009: Scanning electron microscopy studies of antennal sensilla of bruchid beetles, *Callosobruchus chinensis* (L.) and *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae). — *Micron* 40: 320–326.
- Inouchi, J., Shibuya, T., Matsuzaki, O. & Hatanaka, T. 1987: Distribution and fine structure of antennal olfactory sensilla in Japanese dung beetles, *Geotrupes auratus* Mts. (Coleoptera: Geotrupidae) and *Copris pecuarius* Lew. (Coleoptera: Scarabaeidae). — *International Journal of Insect Morphology and Embryology* 16: 177–187.
- Jourdan, H., Barbier, R., Bernard, J. & Ferran, A. 1995: Antennal sensilla and sexual dimorphism of the adult ladybird beetle *Semiadalia undecimnotata* Schn. (Coleoptera: Coccinellidae). — *International Journal of Insect Morphology and Embryology* 24: 307–322.
- Keil, T. A. 1999: Morphology and development of the peripheral olfactory organs. — In: Hansson, B. S. (ed.), *Insect olfaction: 5–47*. Springer Press, Heidelberg. 467 pp.
- Keil, T. A. & Steinbrecht, R. A. 1984: Mechanoreceptive and olfactory sensilla in insects. — In: King, R. C. & Akai, H. (eds.), *Insect ultrastructure: 477–516*. Plenum, New York. 636 pp.
- Kristoffersen, L., Hallberg, E., Wallén, R. & Anderbrant, O. 2006: Sparse sensilla array on *Trioza apicalis* (Homoptera: Triozidae) antennae—an adaptation to high stimulus level. — *Arthropod Structure and Development* 35: 85–92.
- Li, N., Ren, B. Z. & Liu, M. 2007: The study on antennal sensilla of eight Acrididae species (Orthoptera: Acridoidea) in Northeast China. — *Zootaxa* 1544: 59–68.
- Lopes, O., Marques, P. C. & Araujo, J. 2005: The role of antennae in mate recognition in *Phoracantha semipunctata* (Coleoptera: Cerambycidae). — *Journal of Insect Behavior* 18: 243–257.
- Merivee, E. 1992: Antennal sensilla of the female and male elaterid beetle *Agriotes obscurus* L. (Coleoptera: Elateridae). — *Proceedings of the Estonian Academy of Sciences* 41: 189–215.
- Merivee, E., Rahi, M. & Luik, A. 1999: Antennal sensilla of the click beetle, *Melanotus villosus* (Geoffroy) (Coleoptera: Elateridae). — *International Journal of Insect Morphology and Embryology* 28: 41–51.
- Merivee, E., Ploomi, A., Rahi, M., Luik, A. & Sammelse, V. 2001: Antennal sensilla of the ground beetle *Platynus dorsalis* (Pontoppidan, 1763) (Coleoptera: Carabidae). — *Microscopy Research and Technique* 55: 339–349.
- Merivee, E., Ploomi, A., Rahi, M., Bresciani, J., Ravn, H. P., Luik, A. & Sammelse, V. 2002: Antennal sensilla of the ground beetle *Bembidion properans* Steph. (Coleoptera: Carabidae). — *Micron* 33: 429–440.
- Okada, K., Mori, M., Shimazaki, K. & Chuman, T. 1992: Morphological studies on the antennal sensilla of the cigarette beetle, *Lasioderma serricorne* (F.) (Coleoptera: Anobiidae). — *Applied Entomology and Zoology* 27: 269–276.
- Onagbola, E. O., Meyer, W. L., Boina, D. R. & Stelinski, L. L. 2008: Morphological characterization of the antennal sensilla of the Asian citrus psyllid, *Diaphorina citri* Kuwayama (Hemiptera: Psyllidae), with reference to their probable functions. — *Micron* 39: 1184–1191.
- Pophof, B. 1997: Olfactory responses recorded from sensilla coeloconica of the silkworm *Bombyx mori*. — *Physiological Entomology* 22: 239–248.
- Rani, P. U. & Madhavendra, S. S. 1995: Morphology and distribution of antennal sense organs and diversity of mouthpart structures in *Odontopus nigricornis* (Stall) and *Nezara viridula* L. (Hemiptera). — *International*

- Journal of Insect Morphology and Embryology 24: 119–132.
- Rani, P. U. & Madhavendra, S. S. 2005: External morphology of antennal and rostral sensillae in four hemipteran insects and their possible role in host plant selection. — *International Journal of Tropical Insect Science* 25: 198–207.
- Ritcey, G. M. & McIver, S. 1990: External morphology of antennal sensilla of four species of adult flea beetles (Coleoptera: Chrysomelidae: Alticinae). — *International Journal of Insect Morphology and Embryology* 19: 141–153.
- Ruchty, M., Romani, R., Kuebler, L.S., Ruschioni, S., Roces, F., Isidoro, N. & Kleineidam, C. J. 2009: The thermo-sensitive sensilla coeloconica of leaf-cutting ants (*Atta vollenweideri*). — *Arthropod Structure and Development* 38: 195–205.
- Schneider, D. 1964: Insect antennae. — *Annual Review of Entomology* 9: 103–122.
- Silva, C. C., de Capdeville, G., Moraes, M. C., Falcão, R., Solino, L. F., Laumann, R. A., Silva, J. P. & Borges, M. 2010: Morphology, distribution and abundance of antennal sensilla in three stink bug species (Hemiptera: Pentatomidae). — *Micron* 41: 289–300.
- Slifer, E. H. 1970: The structure of arthropod chemoreceptors. — *Annual Review of Entomology* 15: 121–142.
- Smith, C. M., Frazier, J. L., Coons, L. B. & Knight, W. E. 1976: Antennal sensilla of the clover head weevil *Hypera Meles* (F.) (Coleoptera: Curculionidae). — *International Journal of Insect Morphology and Embryology* 5: 349–355.
- Song, L. W., Tao, W. Q., Guan, L., Li, X. P. & Chen, Y. Q. 2010: Influence of host plants and rearing density on growth, development and fecundity of *Arma chinensis*. — *Scientia Silvae Sinicae* 46: 105–110.
- Städler, E. 1984: Contact chemoreception. — In Bell, W. J. & Cardé, R. T. (eds.), *Chemical ecology of insects*: 33–35. Chapman & Hall Press, London. 450 pp.
- Streams, F. A. & Pimentel, D. 1963: Studies on the sex ratio of the harlequin bug, *Murgantia histrionica* (Hemiptera: Pentatomidae). — *Annals of the Entomological Society of America* 56: 712–716.
- Sun, X., Wang, M. Q. & Zhang, G. A. 2011: Ultrastructural Observations on Antennal Sensilla of *Cnaphalocrocis medinalis* (Lepidoptera: Pyralidae). — *Microscopy Research and Technique* 74:113–121.
- Van der Pers, J. N. C. 1981: Comparison of electroantennogram response spectra to plant volatiles in seven species of Yponomeutidae and in the tortricidae *Adoxophyes orana*. — *Entomologia Experimentalis et Applicata* 30: 181–192.
- Ventura, M. U. & Panizzi, A. R. 2005: Morphology of olfactory sensilla and its role in host plant recognition by *Neomegalotomus parvus* (Westwood) (Heteroptera: Alydidae). — *Brazilian Archives of Biology and Technology* 48: 589–597.
- Villela, M. M., Catalá, S., Juberg, J., Silva, I. G. & Dias, J. C. P. 2005: Patterns of antennal sensilla of *Panstrongylus megistus* from three Brazilian states. — *Memórias do Instituto Oswaldo Cruz* 100: 699–702.
- Xu, C. H., Yan, J. J. & Yao, D. F. 1984: The relation between the development of *Arma chinensis* and temperature. — *Scientia Silvae Sinicae* 20: 96–99.
- Xu, C. H., Yao, D. F., Li, Y. M. & Yan, J. J. 1981: A preliminary study of predatory natural enemy, *Arma chinensis* Fallou. — *Practical Forestry Technology* 4: 24–27
- Yu, Q. Q., Liu, Z. K., Chen, C. & Wen, J. B. 2013: Antennal Sensilla of *Eucryptorrhynchus chinensis* (Olivier) and *Eucryptorrhynchus brandti* (Harold) (Coleoptera: Curculionidae). — *Microscopy Research and Technique* 76: 968–978.
- Zacharuk, R. Y. 1980: Ultrastructure and function of insect chemosensilla. — *Annual Review of Entomology* 25: 27–47.
- Zacharuk, R. Y. 1985: Antennae and sensilla. — In: Kerut, G. A. & Gilbert, L. I. (eds.), *Comprehensive insect physiology, biochemistry and pharmacology*: 1–69. Pergamon Press, Oxford. 8356 pp.
- Zahn, D. K., Girling, R. D., McElfresh, J. S., Cardé, R. T. & Millar, J. G. 2008: Biology and reproductive behavior of *Murgantia histrionica* (Heteroptera: Pentatomidae). — *Annals of the Entomological Society of America* 101: 215–228.
- Zou, D. Y., Coudron, T. A., Liu, C. X., Zhang, L. S., Wang, M. Q. & Chen, H. Y. 2013: Nutrigenomics in *Arma chinensis*: transcriptome analysis of *Arma chinensis* fed on artificial diet and chinese oak silk moth *Anthea pernyi* pupae. — *PLoS ONE* 8: e60881.
- Zou, D. Y., Wang, M. Q., Zhang, L. S., Zhang, Y., Zhang, X. J. & Chen, H. Y. 2012: Taxonomic and bionomic notes on *Arma chinensis* (Fallou) (Hemiptera: Pentatomidae: Asopinae). — *Zootaxa* 3382: 41–52.