

A vagrant walrus (*Odobenus rosmarus*) in Finland

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In July 2022, a vagrant female Atlantic walrus (*Odobenus rosmarus rosmarus*) was seen on the south coast of Finland. The same individual, which was called 'Stena' in the international press, had previously been observed in the waters of several other European countries along the North and Baltic Seas. By the time the animal reached Finnish waters it was in poor condition and did not survive a rescue attempt. Post-mortem investigation revealed that the animal was malnourished and its digestive tract was almost empty, but trace amounts of DNA from bivalves and other aquatic invertebrates could be recovered. However, apart from minor age-related ailment and superficial skin wounds, the walrus showed no obvious signs of illness or injuries. Dental wear suggested that the animal was at least 20 years old. Its body and cranial measurements, including tusk length, were well above the average size for a female Atlantic walrus. Mitochondrial DNA supported its origin in the eastern Barents Sea populations. The specimen was mounted and put on display in the Natural History Museum, Helsinki. This is the first confirmed free-ranging walrus observation in the northern part of the Baltic Sea and Finland.

Introduction

The walrus (*Odobenus rosmarus* Linnaeus, 1758) is one of the largest pinnipeds. It has a circum-polar Arctic distribution and the species is divided into two subspecies with geographically distinct ranges: the Atlantic (*O. r. rosmarus*) and

the Pacific (*O. r. divergens* Illiger, 1815) walruses (Fay 1985, Kastelein 2009). The Laptev Sea walrus population, sometimes treated as another subspecies, is now considered to be part of the Pacific taxon on the basis of genetic evidence (Lindqvist et al. 2009, 2016). The Pacific walrus is on average larger than the Atlantic, but

there is considerable overlap in body size (Wiig & Gjertz 1996, Garlich-Miller & Stewart 1998). The Pacific population is currently estimated to comprise more than 200,000 individuals (Beatty et al. 2022), whereas the Atlantic population is 35,000–40,000 walrus, largely because of historical persecution by humans. Atlantic walrus have disappeared from many parts of their former range, such as Canada's Maritime Provinces and Iceland (McLeod et al. 2014, Higdon & Stewart 2018, Keighley et al. 2019, 2022, Garde & Hansen 2021). However, thanks to conservation measures, in recent years walrus have increased in some parts of their distribution area, such as Svalbard (Lydersen et al. 2008, Kovacs et al. 2014). Today, populations of Atlantic walrus are found in north-eastern Canada, Greenland, Svalbard, and the western Russian Arctic including Franz Josef Land and Novaya Zemlya (Wiig et al. 2014, Garde & Hansen 2021).

Vagrant walrus are sometimes seen far south of the species' usual distribution range. In Europe, such sightings are relatively frequent off Norway (Lund 1954, Bruun et al. 1968, Born 1988, Gjertz et al. 1993, Kubny 2022), and in the British Isles (Ritchie 1921, Anonymous 1926, Venables & Venables 1955, Hardy 1959, Cotton 2007, Findlay 2018, Hager 2021, Mullard 2022, Osborne 2023, Wong 2023, Chiacchio & Aae 2024). The southernmost European walrus observations are from southern France and northern Spain (Duguy 1986, Nores & Pérez 1988, Bryant 2021). Other records are from Belgium, the Netherlands, Germany, Denmark, and Sweden (Jensen 1927a, b, Lönnberg 1927, 1940, Redeker 1927, Mohr 1940, Hanström 1943, van Bree 1977, Joensen 1977, Cadée et al. 1982, Reijnders 1982, Mathiasson 1983, Born 1988, Wendehög & Berdenius 2003, Born et al. 2014, Chiacchio & Aae 2024). Because walrus are easy to recognise, conspicuous, and often relatively unbothered by human observers, vagrant individuals attract lots of attention from media and the public, allowing detailed documentation of their movements. Vagrant walrus often also receive affectionate nicknames and cases include 'Wally', who between March and September 2021 visited Ireland, the UK, France, Spain, and Iceland (Bryant 2021, Hager 2021, Mullard 2022); 'Freya', who between October 2021 and August 2022 visit-

ed the Netherlands, Germany, the UK, Denmark, Sweden, and Norway (Fogh-Andersen 2022, Witten 2022b); and 'Thor', who between November 2022 and February 2023 visited the Netherlands, France, the UK, and Iceland (Osborne 2023, Wong 2023).

Initial observations and the walrus' itinerary

On 14 March 2022, an adult walrus was seen on the island of Vigra in Giske, Norway (Google Maps 2022). It received the name 'Stena' by the media in Norway, and although the animal later received other names in other countries, the one given to it originally became the internationally best known. 'Stena's' visit to European waters partially overlapped in time with that of 'Freya', mentioned above. However, these walrus could be identified as different individuals based on unique size and shape of tusks (Witten 2022a). 'Stena' stayed on the Norwegian coast for several weeks, gradually moving southwards (Fig. 1). It was seen in Denmark on 16 May, and on the south-western coast of Sweden on 4 June. On 16 June, the walrus had entered the Baltic Sea, where it was seen off Rügen, Germany. At this stage, the animal appeared to be in good health (Anonymous 2022). The walrus followed the Baltic Sea's eastern coastline and was seen in Poland on 23 June, in Kaliningrad, Russia, on 30 June, and then in Latvia on two occasions. In Latvia, it was first spotted on 3 July on a beach off the city of Liepāja (Pilāts 2022). It was next seen on 7 July, further north in Slītere National Park, where it stayed circa 12 hours lying on the beach. National park rangers observed that the walrus had a fresh small wound of unknown origin in the skin of the lower back.

On 15 July, the walrus came on land by a small boat harbour in Hamina, a town on the southern coast of Finland. The walrus became the object of an extraordinary amount of public interest. The animal's movements were followed closely in the Finnish media, and many news reports and articles were written about it (Valtanen 2023). The walrus remained ashore intermittently in Hamina for almost two days before it returned to the sea and swam away. It next came ashore in the

town of Kotka, circa 18 km west of Hamina, after having briefly become entangled in a fyke net and capsizing a fisherman’s boat. In the morning on 19 July, the walrus was found to have travelled from the top of a narrow bay circa 1.5 km inland along a small watercourse, and then about 50 m by land across suburban backyards, finally ending up in a small patch of forest near a house on private property, where it remained (Karvinen 2022).

Rescue attempt

The walrus was mostly sleeping, and only occasionally adjusted its body position or raised

its head to observe the surroundings (Fig. 2a). It was not observed to defecate. On its ventral side, the animal’s skin had several small superficial wounds, which attracted flies. These wounds probably resulted from its long crawl across dry land, over sharp rocks and various metal objects lying on the ground. At this point, the police had cordoned off the area and only rescue personnel and veterinarians were allowed to approach the animal. An attempt was made to rescue the walrus and transport it to Korkeasaari Zoo in Helsinki. This required anaesthetizing the animal to enable its safe handling and transport. In marine mammals, anaesthetics can trigger a dive reflex that causes life-threatening bradycardia and apnoea (Brunson 2014, Mulcahy & Fravel 2018). In the

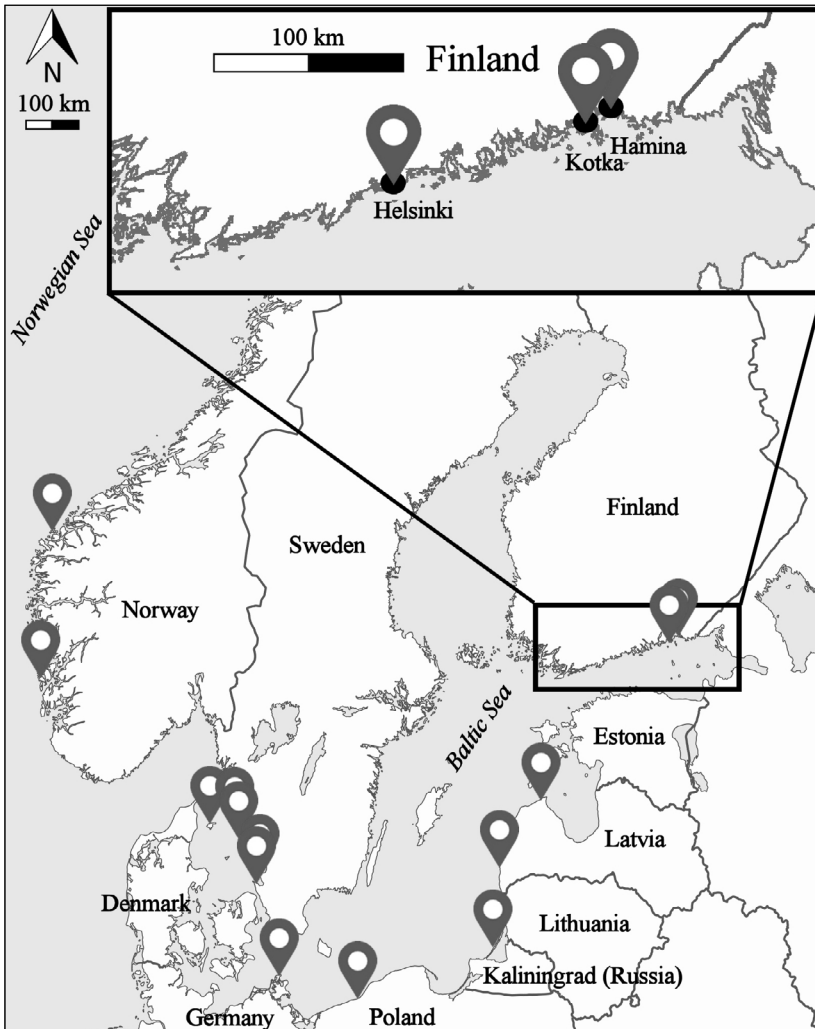


Fig. 1. Map showing the documented itinerary of walrus KX.2531. Base map (modified) from: "Scandinavia location map" by NordNordWest, https://en.m.wikipedia.org/wiki/File:Scandinavia_location_map.svg, CC BY-SA 3.0

present case, the animal was also in a poor physical condition, which increased the risk of anaesthesia-related complications. The animal was lying in a depression in a forest patch and it was estimated that the rescue operation would take hours. Hence, drugs that are suitable for short anaesthesia were not considered (Ølberg et al. 2017).

The animal's size presented an additional challenge. Although the walrus was emaciated, its body mass was still considerable and estimated to be circa 700 kg. The animal was initially sedated with a combination of Zoletil Forte Vet (1:1 tiletamine as hydrochloride and zolazepam as hydrochloride) 1 mg/kg, ketamine (Ketador, Richter Pharma AG, Austria) 2 mg/kg, medetomidine HCl (Medetomidine HCl 30 mg/ml, extem-

poraneously compounded by Yliopiston apteekki, Finland) 0.01 mg/kg, and atropine sulphate (Atropin 1 mg/ml, Takeda Austria GmbH, Austria) 0.03 mg/kg. This combination was a modification of walrus anaesthesia described in Brunson (2014). Drugs were administered intramuscularly into the upper hindquarter with two 20 ml plastic dart syringes with 2 × 60 mm plain needles, which were darted with a CO₂ injection pistol. After the initial dose, the animal got severe apnoea and 300 mg doxapram was hand-injected. This stimulated respiration but also affected the anaesthesia level, making it more superficial. An additional 6.3 mg medetomidine, 500 mg ketamine, and 150 mg Zoletil were hand-injected to secure the safety of the people handling the ani-



Fig. 2. The walrus in Kotka, Finland, on 19 July.

a) The animal resting/sleeping in a forest patch near a suburban house before it was sedated.

b) The eyes of the walrus were covered to keep it calm in case it would awake too soon from anaesthesia. Note the animal's emaciated condition.

Photos: Majju Lanki.



mal. The whole procedure from the first dart to the injection of antidote took approximately 2.5 hours. With the help of a tractor, the animal was lifted with load straps onto a rug; next, it was carried to a transport crate in a lorry (Fig. 2b). Then, 15 mg of atipamezole HCl (Revertor, CP-Pharma Handelsges. mbH, Germany) was hand-injected before the transport to Helsinki started. The walrus however died during transport, shortly after midnight on 20 July.

Post-mortem examination and necropsy

The carcass was taken to the facilities of the Finnish Food Authority in Helsinki for post-mortem analysis. After initial external inspection, it was weighed, measured, and skinned. Next, a necropsy was performed, which included dissection and the taking of samples from all major internal organs. These tissue samples were processed and histologically examined in a routine manner. Bacteriological, virological, parasitological, and faecal DNA samples were taken. In addition to the skin, the bones of the walrus were removed for preservation. The animal's cranium was sawed open during the necropsy to extract the brain.

The necropsy confirmed that the female walrus was starving and in poor body condition. Its subcutaneous blubber layer was very thin and the fat reserves around internal organs and in bone marrow were virtually absent. Although the body dimensions were large (see below), at the time of its death it weighed only 597 kilograms. In ad-

dition to fresh small wounds obtained during the animal's recent overland travels, the skin contained a few old scars. Several specimens of the walrus-specific louse *Antarctophthirus trichechi*, both adults and 3rd instar nymphs (see Scherf 1963, and Leonardi & Palma 2013 for identification), were collected from the skin (Fig. 3). A tick (*Ixodes ricinus*) had also attached itself to the skin while the walrus was ashore. The walrus' lungs were heavily congested and oedematous. The gastrointestinal tract was almost empty, and no macroscopic food material was present in the stomach. Small gastric ulcers were observed in the stomach; these were most likely stress-related. One endoparasitic nematode, the seal worm or cod worm *Pseudoterranova decipiens*, was found in the stomach. In addition, an acute bacterial haemorrhagic cystitis caused by uropathogenic *Escherichia coli* was detected in the urinary bladder. The uterus and ovaries were normal for an old, non-pregnant female mammal. The existence of a corpus albicans but no corpus luteum indicates that the animal had ovulated, and might have been pregnant and given birth during previous reproductive cycles, but not during the current cycle (Garlich-Miller & Stewart 1999). Many organs showed signs of minor age-related ailments, including small benign tumours and unspecific degeneration. Tests for Avian Influenza and SARS-CoV2 virus were negative. No evidence of *Brucella*, *Campylobacter*, *Salmonella*, *Yersinia* or *Leptospira* bacterial infections were detected. Parasitological and coprological examinations for *Toxoplasma gondii*, *Trichinella* sp. and gastrointestinal parasites yielded negative

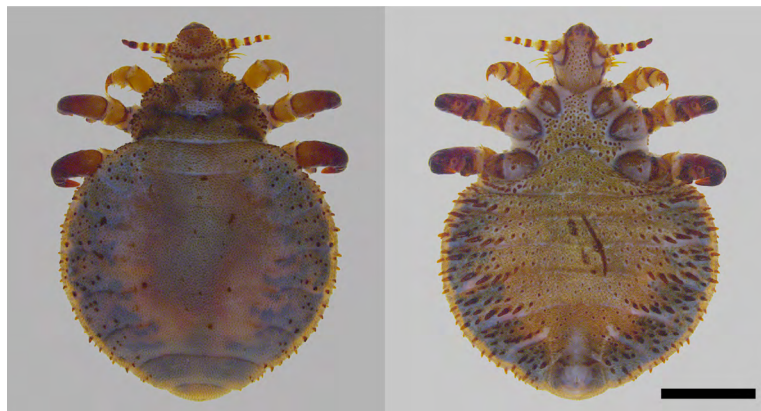


Fig. 3. A female walrus louse *Antarctophthirus trichechi* collected during necropsy. Scale bar 1 mm. Photo: Jaakko Pohjoismäki.

results. Based on necropsy, the cause of death of the walrus was acute cardiac failure, contributed to by emaciated body condition and stress, and most likely triggered by anaesthesia.

NGS analysis of faecal DNA

For information on the diet and on the intestinal microbiome of the walrus specimen, metabarcoding analysis of faecal DNA was performed on two samples, one collected pre-mortem in Latvia, and another collected post-mortem in Finland. The Latvian sample was collected from a beach in Slītere National Park on 8 July, several hours after the animal had left the locality, whereas the Finnish sample was collected from the intestine. The analyses were carried out by Bioname (www.bioname.fi). DNA was extracted by using two optional approaches: with Zymo Research Zymo Quick-DNA Fecal/Soil Microbe Miniprep kit (cat nr: D6010, USA) without toxic beta-mercaptoethanol, and with Macherey-Nagel Nucleo-Spin Stool Kit (ref 740472.250, Germany). The walrus faecal DNA was amplified by using three complementary primer pairs targeting the metazoan mitochondrial *COI* gene: ANML' LCO1490+CO1-CFMRa (Folmer et al. 1994, Jusino et al. 2019), 'fwh' fwhF2+fwhR2n (Vamos et al. 2017), and 'Leray' mlCOLintF+jgHCO2198 (Leray et al. 2013, Geller et al. 2013), one primer pair specifically targeting fish mitochondrial 12S rRNA gene (Teleo-12S-F+Teleo-12S-R; Martínez-Abraín et al. 2020), and one primer pair targeting bacterial 16S rRNA gene (515FB; Parada et al. (2016), modified from Walters et al. (2016); 806RB; Aprill et al. (2015), modified from Caporaso et al. (2011)). Locus-specific PCR setup followed Kankaanpää et al. (2020) and library preparation followed Vesterinen et al. (2016). Next-Generation Sequencing (NGS) was done on partial Illumina NovaSeq6000 SP Flowcell 2x250bp (Illumina Inc., USA) run by the Turku Centre for Biotechnology, Finland. Our bioinformatics pipeline followed Kaunisto et al. (2020).

Only few metazoan taxa were found in the faecal samples in the comprehensive NGS analysis (Table 1). We converted the absolute number of reads in each sample to relative read abundances (RRA; Deagle et al. 2019). In the Latvian sam-

ple, the most abundant taxa were a copepod *Acartia biflosa* (97 % of all the reads), and two marine bivalves, *Kurtiella bidentata* (17.6 % of fwh) and the soft-shell clam *Mya arenaria* (1.9 % of fwh). The sample collected in Latvia also included traces of dipteran insects, presumably a result of contamination from the environment prior to the sampling. The Finnish sample contained traces of a planktonic freshwater cladoceran *Leptodora kindtii* (100 % of the filtered reads for ANML and Leray primer sets, and 96 % for fwh) and a freshwater bivalve, the duck mussel *Anodonta anatina* (3.4 % of the reads). No traces of fish were found in either sample.

The most common bacterial taxa in the Latvian sample were *Lactobacillales* (22 % of the reads), *Psychrobacter* (21 %), *Ignatzschineria* (19 %), *Peptacetobacter* (17 %), and *Glutamibacter* (9 %). The Finnish sample was less diverse and was dominated by *Peptacetobacter* (67 % of the reads) and *Collinsella* (~5.5 %); other microbiont taxa were present with a smaller than 5 % proportion of the reads (Fig. 4).

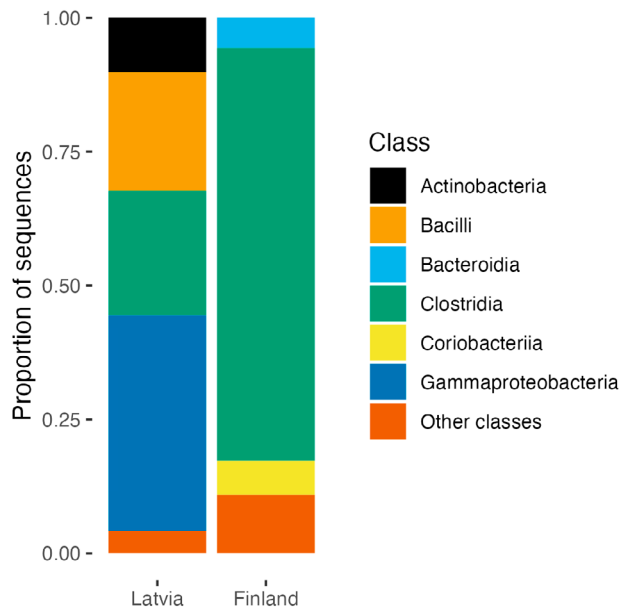


Fig. 4. Comparison of the bacterial composition in faecal samples collected from KX.2531 in Latvia and Finland. Bacterial classes were identified by molecular 16S rRNA analysis. Classes that made up less than 0.05 % of the total proportion are pooled ("Other classes"). ZOTU or genus-wise listing of results is available from author EJV on request.

Table 1. Metabarcoding results of walrus faecal DNA. Metazoan taxa identified in the Latvian and Finnish samples from KX.2531 separately for each of the three primer pairs. 'Count' refers to the number of filtered final sequences assigned to each taxa. 'RRA' refers to relative read abundance, that is, the proportion of sequences assigned to the taxa from the total number of sequences in the sample.

Sample	Phylum	Class	Order	Family	Species	Count	RRA
ANML: LCO1490+CO1-CFMRa							
Latvia	Arthropoda	Copepoda	Calanoida	Acartiidae	<i>Acartia biflosa</i>	40272	0.97
Latvia	Arthropoda	Insecta	Diptera	Anthomyiidae	<i>Fucellia griseola</i>	677	0.02
Latvia	Arthropoda	Insecta	Diptera	Tipulidae	<i>Nephrotoma lundbecki</i>	551	0.01
Latvia	Arthropoda	Copepoda	Calanoida	Acartiidae	<i>Acartia tonsa</i>	86	<0.01
Finland	Arthropoda	Branchiopoda	Haplopoda	Leptodoridae	<i>Leptodora kindtii</i>	6631	1.00
Leray: mIColintF+jgHCO2198							
Latvia	Arthropoda	Copepoda	Calanoida	Acartiidae	<i>Acartia biflosa</i>	14014	0.81
Latvia	Mollusca	Bivalvia	Galeommatida	Lasaeidae	<i>Kurtiella bidentata</i>	3053	0.18
Latvia	Mollusca	Bivalvia	Myida	Myidae	<i>Mya arenaria</i>	326	0.02
Finland	Arthropoda	Branchiopoda	Haplopoda	Leptodoridae	<i>Leptodora kindtii</i>	13921	1.00
fwh: fwhF2+fwhR2n							
Latvia	Arthropoda	Insecta	Diptera	Anthomyiidae	<i>Fucellia griseola</i>	186	0.60
Latvia	Arthropoda	Collembola	Symphyleona	Dicyrtomidae	<i>Dicyrtoma fusca</i>	123	0.40
Finland	Arthropoda	Branchiopoda	Haplopoda	Leptodoridae	<i>Leptodora kindtii</i>	56008	0.97
Finland	Mollusca	Bivalvia	Unionida	Unionidae	<i>Anodonta anatina</i>	2000	0.03

Walrus mitochondrial DNA and specimen origins

To verify the identity and phylogeographic affinities of the walrus specimen, two marker fragments of its mitochondrial genome were sequenced from DNA extracted from muscle tissue: the *COI* gene barcode region (622 bp), and 407 bp of the hypervariable control region (CR), for which extensive reference material is available in public databases (Lindqvist et al. 2008, Andersen et al. 2017). The sequences were deposited in GenBank with the accession numbers OR883405, OR886458, and aligned with available reference data.

The general features of global walrus mtDNA variation include a basic subdivision between the Atlantic and Pacific subspecies, and further subdivision of the Atlantic walrus into an exclusively NW Atlantic clade, and an amphi-Atlantic clade (Lindqvist et al. 2008, Ruiz-Puerta et al. 2023). The *COI* data attributes this walrus specimen to

the latter lineage, but allows for no further resolution. The CR marker is more polymorphic, with 40 different haplotypes in the data of 212 NE Atlantic individuals in Andersen et al. (2017). There are three current subpopulations of the NE Atlantic walrus, those of East Greenland, Svalbard–Franz Josef Land, and the Pechora Sea (including the Kara Sea and Novaya Zemlya), respectively (Garde & Hansen 2021). Of these, the East Greenland subpopulation has a characteristic dominant haplogroup, which is rare elsewhere (haplogroup overlap circa 11 %). This specimen falls within the remaining, predominantly European or Eastern Barents Sea diversity (Fig. 5). It represents haplotype Atl_22, which has previously been recorded both from Franz Josef Land and Pechora Sea walrus specimens (Andersen et al. 2017). Whereas genetic differences between the Svalbard–Franz Josef Land populations and the Pechora Sea populations can be seen when using a larger set of markers, the mitochondrial marker alone cannot separate those provenances.

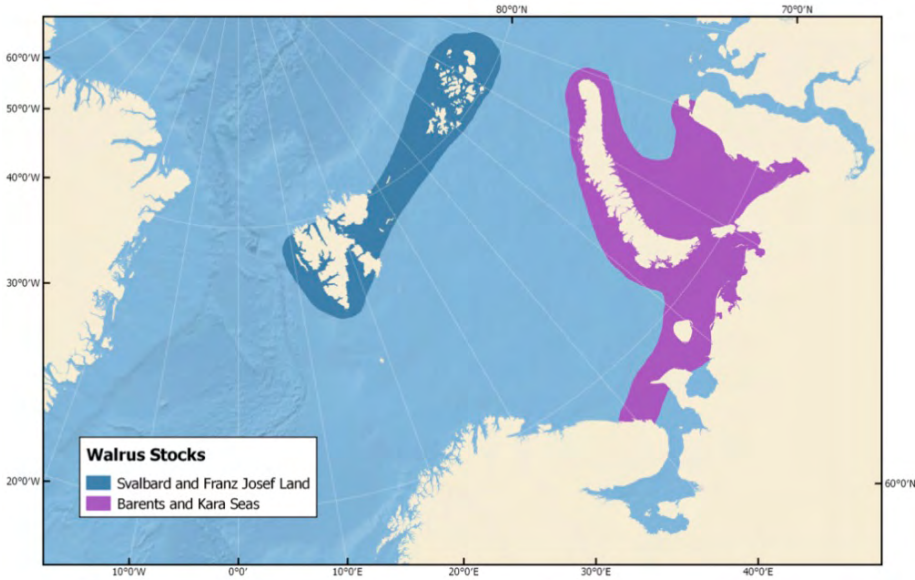


Fig. 5. Distribution of the Svalbard–Franz Josef Land and the Southern Barents Sea (Pechora Sea) and Kara Sea walrus populations. Map by North Atlantic Marine Mammal Commission (NAMMCO); used with permission.

Body and skull measurements

The dorsal curvilinear head–body length of the walrus was 328 cm. The 'standard length' (Scheffer 1967), i.e. the straight-line distance from snout to tip of tail, with the animal lying on its back, was not measured. However, according to Mansfield (1958:107), walrus standard length corresponds to 90 % of the dorsal curvilinear length measurement, and the estimated standard length of the specimen would then be 295 cm. This means that its head–body length was large for a female Atlantic walrus (Table 2). Tail length was not measured from the fresh carcass; however, according to data by Loughrey (1959), tail length in eight female Atlantic walruses was circa 2.67 % of head–body standard length. Thus, the specimen's estimated tail length was circa 7.9 cm.

Several measurements of the skull were taken and compared with measurements published in the literature (Table 2; for definitions of the measurements, see Loughrey 1959, Scheffer 1967, and McLeod et al. 2014). The cranial measurements confirmed that the specimen was large for a female Atlantic walrus. There were no visible injuries in the cranium, but the molariform teeth were heavily worn (Fig. 6). The tusks were long for a female walrus (Fig. 7). Their exposed curvilinear lengths, i.e. measured from the edge of the tooth socket along the length of the outer curve but excluding the root (the "clinical crown" of

Table 2. Body and cranial measurements of the walrus KX.2531 compared with data on female Atlantic walrus from the literature. Weight is in kilograms (kg), other measurements are in centimetres (cm). Tusk length refers to the curvilinear length of the exposed crown. Note that the head–body length and tail length of KX.2531 are estimates; see text for details. Sources: a = Allen 1880; b = Pedersen 1936; c = Mansfield 1958; d = Loughrey 1959; e = Born 1992; f = Knutsen & Born 1994; g = Heptner et al. 1996; h = Garlich-Miller & Stewart 1998.

	KX.2531	range	mean	sources
weight	597	484–808	611	d, f, h
head–body length	295	229–305	267	c, d, e, f, h
tail length	7.90	5.00–9.00	6.10	d
condylobasal length	33.0	23.8–37.3	30.6	a, d, g
zygomatic width	22.3	16.1–25.7	20.4	a, d, g
mastoid width	24.7	19.8–26.2	23.2	a, d
interorbital width	6.89	5.42–7.05	6.06	a, d
rostrum width	15.8	10.0–15.8	13.9	a, d
mandible width	22.8	17.5–23.8	20.9	a, d
tusk length	52.0	8.40–45.0	24.6	a, b, c, d, e, g
upper tooth row	7.42	4.97–7.85	6.26	a, d
lower tooth row	6.55	N/A	N/A	N/A



Fig. 6. Mandible and maxilla of KX.2531, showing molariform tooth rows. Photo: Janne Granroth.



Fig. 7. Lateral view of the skull of KX.2531. Length of metal ruler 30 cm. Photo: Janne Granroth.

Fay 1982:116), were 52.0 cm for the left tusk and 51.5 cm for the right tusk. The total curvilinear lengths of the tusks including the root were 63.6 cm and 62.9 cm, respectively. The tusks showed

relatively little wear, and converged at their tips, which were nearly touching each other; such tusk convergence is typical of female walruses (Allen 1880). No attempt was made to estimate the animal's age based on annual growth layer counts in teeth (Garlich-Miller et al. 1993).

Preservation of the specimen

The Finnish Museum of Natural History decided to preserve the walrus's remains and put the mounted skin on display, whereas the skeleton was to be kept separately in the research collection. The first incision of the skin was made on the right-hand lateral side of the body, as this side would be positioned against the wall and invisible from the viewing side of the mounted animal. To preserve the skin, it was salted, and all muscle and fat tissue were removed. The skin was scoured to remove blood and soluble proteins, shaved to a thickness of circa 1–3 mm, and pickled to acidify it and prevent it from decaying. It was then tanned with Novaltán AL (Zschimmer & Schwarz Chemie GmbH, Germany), and insect-proofed with Eulan SPA 01 (Tanatex Chemicals BV, Netherlands). During the handling of the skin, tatters of epidermis fell off; this was indicative of the fact that the animal had died during the moulting period, which in walruses takes place in the summer (Fay 1985, Kastelein 2009). The enzymatic maceration of the bones with papain (Bauer Handels, Switzerland) followed the pro-

tocol in Niederklopper and Troxler (2001). Bone Maceration Unit MU 1360-2 (Medis Medical Technology GmbH, Germany) was used to carry out the maceration. The bones were degreased with methylene chloride in a Bone Degreasing Unit BDU 1370-100 (Medis Medical Technology GmbH, Germany).

The artificial body, the manikin, was sculpted from polyurethane (Fig. 8a). The soft tissues of the head were modelled onto the skull with oil-based modelling clay (Roma Plastilina, Chavant Clay, USA), and cast in rigid polyurethane foam. The model was fitted with custom-made glass eyes (KL-Glasaugen, Germany) (Fig. 8b). The original tusks were replaced with casts made of PGD polyurethane resin (Creartec Trend-design GmbH, Germany). The skin was fitted onto the manikin and was let dry for three weeks. Patches of skin that had become discoloured during the process were restored by paintbrush with dry acrylic paint. Finally, the 436 whiskers, which had been removed before the processing of the skin began, were each individually re-attached to their original positions (Puolakoski 2024).

The mount with skin (Fig. 9), the skeleton including tusks, frozen muscle tissue samples, as well as DNA extractions thereof, are deposited as preparations in the collections of the Finnish Museum of Natural History, under the stable specimen identifier <http://id.luomus.fi/KX.2531> from which their data are openly accessible.

Discussion

Walrus fossils are known from late Pleistocene deposits on the North Sea coast in Denmark and the Netherlands (Møhl 1974, 1985, Post 2005), but there are no known remains of walrus from the northern Baltic Sea region (Ukkonen 2002, Kangas 2018). Prior to 2022, the only well-documented record from recent times of a vagrant walrus in the Baltic Sea was an adult that visited the coast of north-eastern Germany in April 1939 (Mohr 1940, Hanström 1943). Interestingly, the female that visited Finland in 2022 ('KX.2531' from here on) is strictly speaking not the first walrus ever seen within the present geographical borders of this country. In 1939, a juvenile walrus, originally captured in Greenland, was kept in Helsinki Zoo

for circa three months. In December, this walrus, named 'Turso', managed to escape into the sea off Helsinki. The inexperienced animal presumably perished quickly, possibly by getting lost under the sea ice and drowning; its remains were never found (Lehtonen 1951, Kettunen 2022). Even if it had not escaped, it is unlikely that 'Turso' would have survived very long, as few European zoos were successful in keeping walrus in the early twentieth century (Svanberg 2010).

As noted, KX.2531 was a very large individual. Its estimated head–body length of circa 295 cm approaches the upper limit of published measurements of female Atlantic walrus. While Heptner et al. (1996:32) provided a markedly higher "maximum body length" of 338 cm for female Atlantic walrus, they did not specify whether they referred to curvilinear or standard length (sensu Scheffer 1967). Assuming that it was the curvilinear measurement, using the correction suggested by Mansfield (1958), i.e. 90 %, would result in a maximum standard length of 304.2 cm, which is closer to the estimate for KX.2531. Pedersen (1936) stated that female Greenland walrus reach head–body lengths of "circa 300 cm", similar to the dimension of KX.2531. Unfortunately, Pedersen (1936) specified neither his method of body length measurement nor the number of individuals his data were based on. In contrast, Mansfield (1958), Loughrey (1959), Born (1992), and Knutsen and Born (1994) all measured standard lengths. Mansfield's data set included 29 female Atlantic walrus, Loughrey's 12, Born's 16, and Knutsen and Born's 34. Of these 75 animals, the largest, an individual from Thule, Greenland, in Born's data set, reached 305 cm. The mean female head–body lengths given by these authors are, respectively, 254 cm, 260 cm, 267 cm, and 269 cm. The large size of KX.2531 is also demonstrated by its cranial measurements. In fact, they all exceed the mean values of published measurements (Table 2). However, the sample sizes for published cranial measurements are smaller than for the head–body measurements.

The curvilinear length of the exposed tusks of KX.2531, circa 52 cm, is also notable. In fact, it is longer than all published comparable tusk length measurements of female Atlantic walrus that we know of. Pedersen (1936) provided a tusk length range of 30–35 cm, and Heptner et al.



Fig. 8. Preparation of the mounted specimen. a) Full-body view of the polyurethane manikin. b) Close-up of the head, with the glass eyes in place.

Photos: Ari Puolakoski.



(1996) 27–33 cm. A maximum tusk length of 41.8 cm was reported by Allen (1880) (N = 4), 35.6 cm (N = 27) by Mansfield (1958), 31.4 cm (N = 12) by Loughrey (1959), and 45.0 cm (N = 22) by Born (1992). The tusks of female Pacific walrus are longer. In Fay's (1982) study, the lengths of exposed tusks ("lengths of clinical crown") typically varied between 35 and 55 cm, with a maximum of 68 cm, and the total tusk length was suggested to reach a maximum of 80 cm. Thus, the tusks of KX.2531 were fairly long even by Pacific walrus standards. As the tusks grow through-

out the lifetime, their length also testifies that KX.2531 represented the oldest part of the population, plausibly of 20 years or more; walrus can reach an age of up to 40 years in nature (Taylor et al. 2018, Robeck et al. 2022).

The only measurement which was not unusually large was body weight, undoubtedly due to the emaciated state of the specimen. In Loughrey's (1959) data set of five female Atlantic walrus the maximum body weight was 674 kg and the mean 569 kg, which is slightly less than the weight of KX.2531. Likewise, Knutsen and Born (1994) estimated weights of eight females, with a maximum of 808 kg and a mean of 653 kg. From these data it is clear that at the time of its death, KX.2531 was underweight for its size.

Insights to the reasons for the animal's weight loss may be sought by considering the potential food basis during the journey and evidence about actual diet remains in the metabarcoding of faecal and intestinal samples. Walrus feed primarily on bivalve molluscs and on other benthic macroinvertebrates, of which they may consume tens of kilograms per day (Garda et al. 2018). The bivalve mollusc diversity and available biomass in the brackish Baltic Sea is a fraction of that in marine environments, and the prospects for proper sustenance for the walrus seemed grim at the outset. The first faecal sample from Latvia contained DNA traces of two bivalves. *Mya arenaria* is the largest of the marine bivalves in the Baltic (> 5

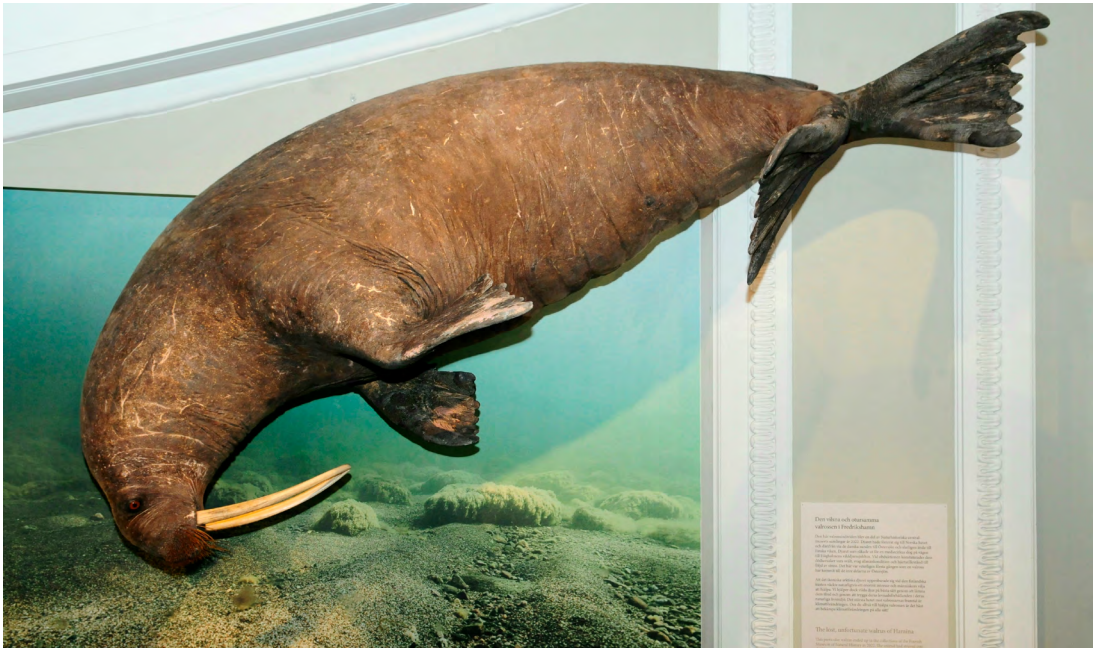


Fig. 9. The finished mounted specimen, prepared by Chief Taxidermist Ari Puolakoski, on display in the Natural History Museum, Helsinki. Photo: Janne Granroth.

cm) and perhaps the most likely prey item there. *Kurtiella bidentata* (= *Mysella bidentata*) however is a tiny clam (< 1 cm) and an unlikely prey item. Also, its distribution does not extend to the inner Baltic (e.g. Morys et al. 2017); if the walrus actually proceeded consistently east and north, it hardly could have ingested *Kurtiella* within two-three weeks before it was seen in Latvia.

The benthic biomass in the Eastern Gulf of Finland where the walrus was last encountered, with salinity < 5, is very low, c. 20 g/m² (Gogina et al. 2016). In the offshore zone, which might resemble the walrus' normal feeding habitat, the Baltic clam *Macoma balthica*, of a size 1–2 cm, is practically the only available bivalve, characterizing the macrozoobenthos along with *Marenzelleria* polychaetes. On the other hand, distributions of many freshwater species extend to the lowest-salinity nearshore waters of the Baltic. One of these species is the unionid mussel *Anodonta anatina*, which is of a proper prey size, up to 10 cm length, and accounts for a specific habitat type in river mouths, inlets and inshore lagoons, "benthic habitats characterized by Unionidae" ("suursimpukkapohjat"; Kotilainen et al. 2020). The two landing places of the walrus were

in the immediate vicinity of documented unionid habitats (Velmu 2024; Ari Laine, personal communication), in the very innermost bays, each several kilometres long and isolated by narrow straits at their entrance. It initially seemed possible that these inshore explorations were affected by cues of mollusc prey in the unfamiliar environment. With the first stranding in Hamina, the walrus defecated prominently, suggesting it still had foraged on the way. Three days later its intestinal tract was empty of food remains, but a trace of *Anodonta* DNA was recovered, indicating that it at least had attempted to feed on the freshwater mussel.

The differences between the bacterial composition of the Latvian and Finnish faecal samples suggest that the animal's gut microbial diversity had become diminished over time. This is likely due to the walrus' poor diet during its travel in the Baltic Sea.

The failure of the attempted rescue operation of the walrus was tragic but not unexpected. Because of their size and potential aggressiveness, walrus are difficult to handle and transport. To our knowledge, there has been only one successful rescue and release operation of a vagrant wal-

rus in European waters. An approximately five-year old male walrus of circa 400 kg was captured with a net, without the use of anaesthesia, in the Netherlands in 1981; it was first transported by truck and ferry to a temporary holding site for observation, and later released into the North Sea (Reijnders 1982). This walrus was not only younger and smaller but also apparently in better physical condition than KX.2531.

From its place of origin in the eastern Barents Sea, KX.2531 must have travelled at least 3,000 km to reach the southern coast of Finland. Generally, young animals are more likely to travel long distances in search of a new territory or a mate than older animals are. In most marine mammals, males typically disperse longer distances than females, and this also holds for the walrus (Li & Kokko 2019; Mesnik & Ralls 2009). Tagging experiments have shown that male walrus may undertake movements of up to 1,500 km to reach new haul-out sites (Semenova et al. 2019). With such behaviour young males would also be more likely to become lost and end up far from their natural range. Several of the vagrant walrus in Europe have indeed been young males (e.g. Joensen 1977, Reijnders 1982, Bryant 2021, Mullard 2022). However, KX.2531 was a mature female that may have been past its reproductive prime.

Possible explanations for vagrant walrus may be sought in a combination of increased population size and environmental change. In the eastern parts of the Atlantic range, including the Russian Arctic, walrus have been legally protected since the 1950s, which has led to population recovery (Gjertz et al. 1998, Lydersen et al. 2012, Kovacs et al. 2014, Wiig et al. 2014, Semenova et al. 2019, Norwegian Polar Institute 2022). Population growth and ensuing increased competition for space and other resources could induce unusually long individual dispersal distances (Born 1988, Born et al. 2014, 2021). While hunting pressure has ceased, walrus have simultaneously had to contend with environmental change. The recent climatic changes are having an especially strong effect in Arctic regions by reducing the sea ice cover (Rantanen et al. 2022). The observed and projected sea ice loss is particularly extensive in the Barents Sea (Rieke et al. 2023). Sea ice is important for walrus, which

must leave the water during periods of resting, moulting, and reproduction (Kovacs et al. 2015, Born et al. 2021). Sea ice reduction may also affect walrus foraging. Walrus feed mainly on benthic invertebrates, and their foraging is thus limited by water depth. Although walrus are capable of diving deeper, their daily foraging dives are typically concentrated to depths of less than 100 metres (Garde et al. 2018). Thus, foraging walrus tend to remain in coastal waters, within distances of 40–100 km from their haul-out sites (Garde & Hansen 2021). As a haul-out substrate, sea ice allows walrus to reach foraging areas further away from the coast. Although walrus can utilise both dry land and ice as haul-out substrates, a decrease of sea ice cover may lead to a net loss of suitable haul-out sites (Jay et al. 2012). It has been suggested that Atlantic walrus may be less vulnerable to such effects than Pacific walrus, as the current population size of the Atlantic subspecies is lower relative to its carrying capacity, and because the continental shelf is narrower in the Atlantic distribution area and feeding habitat is thus accessible also from land-based haul-outs (Born et al. 2021).

However, although there has been a general decreasing trend in sea ice cover in the Barents Sea in the period 1979–2023, it should be noted that the winter of 2021–2022 was not especially mild. In January and February 2022, the months immediately preceding the first observations of KX.2531 off the coast of Norway, the extent of ice cover in the Barents Sea was larger than it has been in many recent years (Norwegian Ice Service 2023). On the other hand, historical records suggest that long before the onset of anthropogenic climate change, vagrant walrus have been observed off the coasts of Europe and the Atlantic coast of North America (e.g. Ritchie 1921, Allen 1930, Löwegren 1944, Kiparsky 1952, Lund 1954, Manville & Favour 1960, Cotton 2007, Monaghan 2017, Mullard 2022, Chiacchio & Aae 2024). Thus, the scientific evidence for environmental change as a driver of Atlantic walrus extralimital movements is not clear-cut, and it cannot be stated with any certainty that the occurrence of KX.2531 in the Baltic Sea is related to climate change in general. The ultimate causes behind this individual's unusual visit to Finland remain unknown.

Acknowledgements. We would like to thank **Carsten Grøndahl** and **Rolf-Arne Ølberg** for support and advice with the anaesthesia plans, and **Jenni Hyväkkä** and **Tuire Tuukkanen** for co-operation during the rescue attempt. We also thank **Heini Nihtilä** and **Nina Trontti** for assistance and organizing logistics. We thank **Pasi Sihvonen**, **Sanna Vuori** and **Hanna Laakkonen** for managing the acquisition of the walrus to the Natural History Museum, Helsinki, **Elina Laiho** for DNA laboratory work, and **Roni Andersson**, **Heikki Ahopelto** and **Elina Laurén** for their contributions to the preparation of the mounted specimen. The walrus faecal DNA samples from Slitere National Park, Latvia, were collected by **Vilnis Skuja**, **Madars Burnevics** and **Gunita Dekšne**. Special thanks to **Mirka Hlavaty** for helping with the NGS analysis. We wish to acknowledge the DNA analysis company **Bioname** for their expertise and help regarding the molecular analysis and interpretation of the results. We thank the **Finnish Functional Genomics Centre** supported by University of Turku, Åbo Akademi University, and Biocenter Finland. **Mervi Kunnasranta** and **Marja Isomursu** provided valuable comments which improved the manuscript.

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